

New Natural Weed Hosts of *Sweet potato feathery mottle virus* in SyriaEnsaf Akel¹, Imad D. Ismail¹, Salah Al-Chaab² and Segundo Fuentes³

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Abstract

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A survey was conducted along the Syrian coastal area to identify natural weed hosts of *Sweet potato feathery mottle virus* (SPFMV, genus *Potyvirus*, family *Potyviriidae*). Eight hundred sixty eight selected samples representing fifty six species (forty five genera, twenty three families) were collected from ten fields related to five regions (Zagrin, Sarsakia, Brgan, Ras Al-Aen, and Hreson) during 2006/2007 and 2007/2008 growing seasons. All samples were tested for the presence of *Sweet potato feathery mottle virus* by using tissue blot immunoassay and polyclonal antibodies. Results showed that nineteen species were naturally infected with *Sweet potato feathery mottle virus* including *Amaranthus retroflexus* L., *Atriplex hastata* L., *Brasica oleracea* var. capitata., *Calendula officinalis* L., *Capsicum annum* L., *Chenopodium* sp., *Chenopodium hybridum* L., *Chenopodium polyspermum* L., *Chenopodium vulvaria* L., *Convolvulus arvensis* L., *Cucurbit maxima* Dush., *Heliotropium europaeum* L., *Ipomoea violacea* L., *Malva neglecta* Wallr., *Mentha arvensis* L., *Phaseolus lanatus* L., *Sinapis arvensis* L., *Spinacea oleracea* L and *Traxacum officinale* Web belonging to twelve families and mostly to the families *Chenopodiaceae* and *Convolvulaceae*. All weed species were recorded for the first time in Syria as natural hosts of SPFMV.

Keywords: SPFMV, sweet potato, Syria, viruses, weed.

Introduction

Volunteer plants and weeds provide shelter and sources of nutrients for virus vectors. Other vegetative structures or contaminated weed seeds may also harbor viruses. Aside from facilitating the spread of disease as alternative source of inoculum, these plants sustain the viability of the virus between crop seasons (16). Numerous weeds that act as reservoirs for a virus and its vectors (6, 22), and certainly those of genus *Datura*, have been found to be important in the epidemiology of certain viral diseases (17, 20, 25). *Sweet potato feathery mottle virus* (SPFMV; genus *Potyvirus*; family *Potyviriidae*) is the most common virus on sweet potatoes (*Ipomoea batatas* (L.) Lam) in Africa (24, 32, 33, 38) and elsewhere in the world (7, 9, 11, 26, 28, 30, 35) and causing variable symptoms and damage to the crop (11, 26, 28, 30). The first report of a SPFMV incidence was from Africa (15). Later it was reported from Italy (32), and recently in the Syrian coastal area (2, 21). SPFMV is transmitted by aphids, *Aphis gossypii*, *A. craccivora*, *Lipaphis erysimi*, *Myzus persicae*, in a nonpersistent manner (4, 5, 13, 18, 35). The virus can also be transmitted mechanically, and by grafting (2, 4, 8, 29). It is not transmitted by contact between plants, by seed, or by pollen (7). Many isolates and strains of SPFMV have been characterized in different parts of the world. Some strains of SPFMV cause economic losses by their effect on storage root quality (internal cork and russet crack) (19, 29). In the last few years, four strains of SPFMV (RC, O, C and EA) have been reported (34, 36). Two strains were found in North Carolina (31). Four strains of SPFMV were recorded in Syria based on their reaction on the indicator plants *I. setosa* and *I. nil* (3). In Japan, most of sweet potato cultivars were susceptible to a SPFMV-S strain, which causes serious production losses (39). An East African strain of SPFMV incited severe 'sweetpotato virus disease' in plants

co-infected with *Sweet potato chlorotic stunt virus* and threatens subsistence sweetpotato production. However, little was known about its natural hosts and ecology, although SPFMV incidence was similar in annual and perennial species (37). The experimental host range of the virus is mainly restricted to the *Convolvulaceae* and *Chenopodiaceae* families, but a few SPFMV strains also infect species of the family *Solanaceae* of which *Nicotiana benthamiana* is a good propagation host for virus purification (12).

In Louisiana, United states, the perennial wild plants of *I. trichocarpa* Ell. and some annual wild species such as *I. hederacea* Jacq., *I. hederifolia* L., *I. lacunose* L., and *I. wightii* (Wall) were infected with SPFMV in the field and may act as a natural reservoir of SPFMV which can then be transmitted to sweet potato (12). In East Africa, over 89 species of *Ipomoea* (40) and several species of other genera of Convolvulaceae were known to occur, including one species each of the genera *Hewittia* (Wight & Arn) and *Lepistemon* (Blume) (40). In central Uganda, *I. eriocarpa* R.Br., *I. hederifolia* L., and *I. tenuirostris* Chisy were reported to be infected with SPFMV (23, 24).

The main objective of this research is to identify the weeds which can host SPFMV under Syrian conditions.

Materials and Methods

Field Survey

Eight hundred and sixty eight selected foliar samples representing 56 weed species, 45 genera, and 23 families were collected from 10 sweet potato fields and from the surroundings area related to 5 regions covering Zagrin, Sarsakia, Brgan, Ras Al-Aen, and Hreson in Lattakia and Tartous governorates, during 2006/2007 and 2007/2008 growing seasons. Regions were selected on the basis of high incidence of sweet potato plants exhibiting

characteristic symptoms of SPFMV. Samples were blotted on nitrocellulose membranes according to Makkouk and Kumari (27).

Detection of SPFMV

Tissue blot immunoassay (TBIA) was used for serological detection of SPFMV with polyclonal antibodies provided by the International Potato Center (CIP), Lima, Peru (10). The kit also contained NCM strips spotted with sap from SPFMV-positive and negative controls. Samples were also spotted on nitrocellulose membranes in Syria and processed at CIP to confirm results.

Results and Discussion

A large number of weed species with symptoms suggestive of virus infection were observed in sweet potato fields and their surroundings in Syrian coastal area during 2006/2007 and 2007/2008 growing seasons from April to October (Table 1). *Amaranthus retroflexus* L., *Convolvulus arvensis* L., *Chenopodium* sp. were common weed species in most surveyed fields. Mosaic and leaf malformation symptoms were the most common. Some collected samples showed mosaic, mottle, yellowing, chlorosis, stunting and necrotic spots, whereas other samples did not show any obvious symptoms.

Results of TBIA test showed that 19 weed species were infected with SPFMV (Table 1). SPFMV-infected wild host plants identified in this study belong to different families and not only to *Convolvulaceae*. Individual SPFMV-infected wild plant could serve as a virus reservoir for many years, encompassing several sweet potato growing seasons. In Louisiana, the perennial weed *I. trichocarpa* is reported as a reservoir of SPFMV throughout sweet potato growing seasons (12). However, there are no studies to show whether isolates of SPFMV isolated from wild species play a role in the incidence of SPFMV in surrounding sweet potato fields. There is no similar study in literature aimed to survey the host species of SPFMV under natural infection conditions, but some previous studies aimed to search host range under artificial infection conditions (1, 8, 14). A study done by Tugume *et al.* (37) indicated that SPFMV was prevalent at different incidences in 22 *Ipomoea* species, and in *Hewittia sublobata*, *Lepistemon owariensis*, of which 19 species were new natural hosts for SPFMV. *I. eriocarpa*, *I. hederifolia*, *I. wightii*, and *I. tenuirostris*, previously were known as natural hosts of SPFMV in Uganda (23, 24). *I. hederifolia* and *I. wightii* were found infected with SPFMV also in Louisiana (12).

New sweet potato cultivations are established by taking cutting from vines of the previous crops for planting, which makes sweet potato a perennial plant in the local cropping system. Although farmers usually manage to select symptomless and SPFMV-free vines as source of cuttings (18), the subsistence sweet potato crops and especially the remaining plants from harvested crops and those grown in abandoned plots often are found infected with SPFMV (18). The actual interactions of the weed and wild species and sweet potato crops in relation to virus transmission require further study.

Table 1. Weed and natural hosts of SPFMV recorded in Syria. in Sweet potato fields and their surroundings in 2006/2007 and 2007/2008 growing seasons. Letter with underlines refer to symptoms gives positive reaction with SPFMV antiserum

Family Weed species	External symptoms	No. of infected samples/ No. of tested samples
Amaranthaceae		
<i>Amaranthus hybridus</i> L.	SNS	0/4
<i>A. retroflexus</i> L.	Mm, Vby, <u>M+Mo</u> , <u>Nsy</u> , Y, Ch, Vby +YSS, St+SNS, MO, <u>St+Vby</u> , <u>My+Ldi</u> , My, <u>Mm</u>	16/172
Apiaceae		
<i>Scandixpecten-veneris</i> L.	M	0/8
Asteraceae		
<i>Calendula officinalis</i> L.	<u>YSS</u> , Ch	1/8
<i>Xanthium strumarium</i> L.	My	0/4
Boraginaceae		
<i>Heliotropium europaeum</i> L.	<u>Nsy</u>	0/28
<i>Anchus officinalis</i> L.	Nsy	0/12
Brassicaceae		
<i>Brasica oleracea</i> var Capitata	<u>Vby</u>	0/4
<i>Raphanus raphanistrum</i> L.	My, Mo	0/8
<i>Sinapis arvensis</i> L.	<u>Nsy</u> , Mo	0/8
Caryophyllaceae		
<i>Cerastium arvense</i> L.	Vby	0/12
Chenopodiaceae		
<i>Atriplex hastate</i> L.	<u>Ldi</u>	1/4
<i>Chenopodium</i> sp.	My, <u>Nsy</u> , YSS	4/32
<i>C. amaranticolor</i> L.	Nsy	0/24
<i>C. quinoa</i> Willd.	Nsy	0/8
<i>C. hybridum</i> L.	<u>Nsy</u>	1/4
<i>C. polyspermum</i> L.	<u>Nsy</u>	1/4
<i>C. vulvaria</i> L.	<u>M</u>	1/4
<i>Spinacea oleracea</i> L.	<u>YSS</u> , Y, Mm, Nsy	2/16
Compositae		
<i>Artemisia vulgaris</i> L.	Nsy	0/4
<i>Bidens pilosa</i> L.	SNS, Nsy, Mo	0/16
<i>Lactuca sativa</i> L.	M+Ldi, Y	0/8
<i>Sonchus oleraceus</i> L.	YSS, Nsy	0/16
<i>Traxacum officinale</i> Web.	<u>M</u> , Nsy, Ch, Mo, Y	2/32
Convolvulaceae		
<i>Convolvulus arvensis</i> L.	SNS, YSS, Mo, Vby, St, M, <u>Nsy</u>	8/124
<i>Ipomoea violacea</i> L.	<u>Lc+Vy</u> , Ldi+Vy	2/8
Cucurbitaceae		
<i>Cucurbita maximx</i> Dush.	<u>Mm</u>	1/8
<i>C. pepo</i> L.	Mo	0/4
<i>Cucumis melo</i> L.	M	0/4
<i>C. sativus</i> L.	M	0/8
<i>Luffa cylindrical</i> L.	M	0/8
<i>Citrullus vulgaris</i> L. (thunb)	Nsy, Ch	0/8
Euphorbiaceae		
<i>Euphorbia heterophylla</i> L.	Vby, Nsy, SNS	0/24
<i>E. helioscopia</i> L.	Nsy	0/8
<i>E. peplus</i> L.	Nsy	0/4
Graminae		
<i>Digitaria sanguinalis</i> L.	Nsy	0/16
<i>Synodon dactylon</i> pers	Nsy	0/8

Table 1 (Cont.)

Family Weed species	External symptoms	No. of infected samples/ No. of tested samples
Labiatae		
<i>Mentha arvensis</i> L.	Ldi	1/4
Leguminaceae		
<i>Phaseolus lanatus</i> L.	M+Ldi, M, Y	1/12
<i>Ph. vulgaris</i> L.	M, Ldi	0/8
Malvaceae		
<i>Althaea officinales</i> L.	My	0/8
<i>Hibiscus trionum</i> L.	Nsy	0/4
<i>Malva neglecta</i> Wallr.	Y, Vby, St, Nsy	4/28
Papaveraceae		
<i>Papaver rhoeas</i> L.	Nsy	0/4
Pedaliaceae		
<i>Sesamum indicum</i> L.	Mo	0/4
Polygalaceae		
<i>Rumix crispus</i> L.	SNS	0/4
Portulacaceae		
<i>Portulaca oleracea</i> L.	Nsy	0/8
Primulaceae		
<i>Anagallis arvensis</i> L.	Nsy	0/4
Solanaceae		
<i>Capsicum annuum</i> L.	M+Ldi, MO, M	4/16
<i>Datura stramonium</i> L.	M.	0/4
<i>Hyoscyamus niger</i> L.	Nsy	0/4
<i>Lycopersicon esculentum</i> Miller	St+My, St+Ldi, Y, Mo, Vby, St, M	0/36
<i>Nicotiana tabacum</i> L.	M, Nsy, My, YSS	0/12
<i>Solanum nigrum</i> L.	Mm, Mo, Nsy	0/20
Sycophulariaceae		
<i>Veronica persica</i> Poir.	Nsy	0/4
Tiliceae		
<i>Corchorus olitorius</i> L.	Vby, Mo, YSS	0/12

* Ldi= leaf malformation; Nsy= No-symptoms; Mo= mottling; SNS= Necroses spot; M= Mosaic; MMO= Mild mosaic; Vby= Yellow between vein; Vy= Vein-clearing; YSS= Yellow small spot; Y= Yellow; My= Yellowing Mild; St= Stunting; Ch= Chloroses; Lc= Leaf curl; Mm= Mild mottling.

All plants in the fields are subject to attack by numerous pests (11, 28). In this regard, the plants/samples which showed symptoms and gave positive reaction with SPFMV are listed in Table 1. The observed symptoms are not necessarily caused only by SPFMV infection. On the other hand, all plants/samples which exhibited external symptoms and gave negative reaction with SPFM, cannot be considered as non-hosts for the virus. It could well be that at the time of the survey, such species were not exposed to SPFMV infection. In addition, such hosts can be favorable to the insect vectors. *Chenopodium amaranticolor* L and *C. quinoa* Willd were SPFMV free under natural infection conditions (Table 1) but they are reported as SPFMV host under laboratory conditions (1, 8, 14). To confirm our results, weed species which did not react positively for SPFMV presence in TBIA test should be tested again under artificial inoculation conditions and then tested serologically. Perennial plant species infected with SPFMV were found more important as alternative hosts of the virus, because its ability to play the role of source of inoculum, for the neighbouring plants in the field or other neighbouring fields, for many years.

SPFMV is not known to be seed transmitted in sweet potato. Presuming that this also holds true for the wild species, a hypothesis which needs to further investigation.

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المخلص

عاقل، إنصاف حسن، عماد اسماعيل، صلاح الشعبي، وسيجوندو فوينتس. 2010. العوامل العشبية الطبيعية لفيروس التبرقش الريشي للبطاطا الحلوة في سورية. مجلة وقاية النبات العربية، 28: 96-100.

أجري مسح حقلي للتعرف على العوامل العشبية الطبيعية لفيروس التبرقش الريشي للبطاطا الحلوة في الساحل السوري. جمع خلاله 868 عينة نباتية فردية من 56 نوع نباتي (45 جنساً، و23 عائلة) وذلك من 10 حقول عائدة لـ 5 مناطق رئيسية لزراعة البطاطا الحلوة (زغرين- السرسكية- البرجان- رأس العين- حريصون)، خلال موسمي 2007/2006 و2008/2007. اختبرت جميع العينات باستخدام بصمة النسيج النباتي المناعية والمصل المضاد لفيروس التبرقش الريشي متعدد الكلون. أكدت النتائج الإصابة الطبيعية بفيروس التبرقش الريشي لـ 19 نوع من العينات العشبية والمحصولية المختبرة (*Amaranthus* L., *retroflexus* L., *Atriplex hastata* L., *Brasica oleracea* var. capitata., *Calendula officinalis* L., *Capsicum annuum* L., *Chenopodium sp.*, *Chenopodium hybridum* L., *Chenopodium polyspermum* L., *Chenopodium vulvaria* L., *Convolvulus arvensis* L., *Cucurbita maxima* Dush., *Sinapis arvensis* L., *Phaseolus lanatus* L., *Mentha arvensis* L., *Malva neglecta* Wallr., *Ipomoea violacea* L., *Heliotropium europaeum* L., *Spinacea oleracea* L. و *Traxacum officinale* Web.)، وهذه الأنواع عائدة لـ 12 عائلة نباتية مختلفة، وقد وجد أن معظم العينات المصابة بالفيروس تنتمي إلى عائلتي *Convolvulaceae* و *Chenopodiaceae*، وبعد اكتشاف هذه الأنواع النباتية المصابة بفيروس التبرقش الريشي التسجيل الأول للإصابة الطبيعية في سورية باستثناء البطاطا الحلوة العائل الرئيس لهذا الفيروس.

كلمات مفتاحية: SPFMV، بطاطا حلوة، سورية، فيروسات، أعشاب.

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References

1. Akel, E.H. 2005. A survey of some sweet potato virus diseases in Syrian coast. MSc thesis. Plant Protection, Faculty of Agriculture, Tishreen University. Syria. 119 pp.
2. Akel, E.H., I.D. Ismail and S. Raee. 2006. Diagnosis of some sweet potato viruses by using indicator plants and serological tests. Tishreen University Journal, 28: 161- 173.
3. Akel, E.H., I.D. Ismail and SH.M. Salah. 2008. Identification of some Sweet potato feathery mottle Potyvirus isolates by using differential indicator plants Tishreen University Journal (in press)
4. Aritua, V., E. Adipala, E.E. Carey and R.W. Gibson. 1998a. Aspects of resistance to sweet potato virus disease in sweet potato. Annals of Applied Biology, 132: 387-398.
5. Aritua, V., E. Adipala, E.E. Carey and R.W. Gibson. 1998b. The incidence of sweet potato virus disease and virus resistance of sweet potato grown in Uganda. Annals of Applied Biology, 132: 399-411.
6. Bock, K.R. 1982. Geminivirus diseases in tropical crops. Plant Disease, 66: 266- 70.
7. Brunt, A.A., K. Crabtree, M.J. Dallwitz, A.J. Gibbs and B. Watson. 1996. Viruses of Plants. Descriptions and Lists from the VIDE Database. CAB International, Wallingford, U.K.
8. Cali, B.B. and J.W. Moyer. 1981. Purification, serology, and particle morphology of two russet crack strain of sweet potato feathery mottle virus. Phytopathology, 71: 302-305.
9. Cadena-Hinojosa, M.A. and R.N. Campbell. 1981. Characterization of isolates of four aphid-transmitted sweet potato viruses. Phytopathology, 71: 1086-1089.
10. CIP (International Potato Center). 2001. Techniques in plant Virology in CIP Training Manual Version, 10 January 2001. L. F. Salazar and U. Jayasinghe (eds.). CIP, Lima, Peru.
11. Clark, CA. and J.W. Moyer. 1988. Compendium of Sweet Potato Diseases. The American Phytopathological Society, St. Paul, MN.
12. Clark, C.A., K.S. Derrick, C.S. Pace and B. Watson. 1986. Survey of wild *Ipomoea* species as potential reservoirs of sweet potato feathery mottle virus in Louisiana. Plant Disease, 70: 931-932.
13. Cucho, F. 1993. Distribution of main sweet potato (*Ipomoea batatas* (L.) Lam) viruses in the south of Peru (NAZCA-Canete). Sanluis Gonzaga National university. Agronomic Eng. Thesis. Ica, Peru. 86 pp. (In Spanish).
14. Difeo, L., S.F. Nome and E. Biderbost. 2000. Etiology of sweet potato chlorotic dwarf disease in Argentina. Plant Disease, 84: 35-39.
15. Doolittle, S.P. and L.L. Harter. 1945. A graft-transmissible virus of sweet potato. Phytopathology, 35: 695-704.
16. Duffus, J. 1971. Role of weeds in the incidence of virus diseases. Annual Review of Phytopathology, 9: 319-340.
17. Fereres, A., C. Avila, J.L. Collar, M. Duque and C. Fernandez-Quintanilla. 1996. Impact of various yield-reducing agents on open-field sweet peppers. Environmental Entomology, 25: 983-986.
18. Gibson, R.W., I. Mpenbe, T. Alicai, E.E. Carey, R.O.M. Mwanga, S.E. Seal H.J. and Vetten. 1998. Symptoms, etiology and serological analysis of sweet potato virus disease in Uganda. Plant Pathology, 47: 95-102.
19. Gibson, R.W., R.O.M. Mwanga, S. Kasule, I. Mpenbe and E.E. Carey. 1997. Apparent absence of viruses in most symptomsless field-grown sweet potato in Uganda. Annals of Applied Biology, 130: 481-490.
20. Hobbs, H.A., D.M. Eastburn, C.J.D. Arcy, J.D. Kindhart, J.B. Masiunas and D.J. Voegtlin. 2000. Solanaceous weeds as possible sources of Cucumber mosaic virus in Southern Illinois for aphid transmission to pepper. Plant Disease, 84:1221-1224.
21. Ismail, I.D., S. Raee and E.H. Akel. 2004. A survey of sweet potato virus diseases in Syrian coastal region "Lattakia" By using Tissue blot immunoassay. Tishreen University Journal, 26: 161-179.
22. Kahn, N.D., J.F. Walgenbach and G.C. Kennedy. 2005. Summer weeds as hosts for *Frankliniella occidentalis* and *Frankliniella fusca* (Thysanoptera: Thripidae) as reservoirs for Tomato spotted wilt tospovirus in North Carolina. Journal of Economic Entomology, 98:1810-1815.
23. Karyeija, R.F., R.W. Gibson and J.P.T. Valkonen. 1998a. Resistance to sweet potato virus disease (SPVD) in wild East African *Ipomoea* spp. Annals of Applied Biology, 133: 39-44.
24. Karyeija, R.F., R.W. Gibson and J.P.T. Valkonen. 1998b. The significance of sweet potato feathery mottle virus in subsistence sweet-potato production in Africa. Plant Disease, 82: 4-15.
25. Latham, L.J. and R.A.C. Jones. 1997. Occurrence of Tomato spotted wilt tospovirus in native flora, weeds, and horticultural crops. Australian Journal of Agricultural Research, 48: 359-369.
26. Lopez, D. and L.F. Salazar. 1987. Studies on sweet potato feathery mottle virus (SPFMV) in Peru. Fitopatologia 22: 40-41 (Abstract in Spanish).
27. Makkouk, K.M. and S.G. Kumari. 1996. Detection of ten viruses affected legume crops by tissue blot immunoassay (TBIA). Arab Journal of Plant Protection, 141: 3-9.
28. Moyer, J.W. and L.F. Salazar. 1987. Viruses and virus-like diseases of sweet potato. Plant Disease, 73: 451- 455.
29. Moyer, J.W. and G.G. Kennedy. 1978. Purification and properties of sweet potato feathery mottle virus. Phytopathology, 68: 998 -1004.
30. Moyer, J.W. 1986. Variability among strains of SPFMV. Phytopathology, 76:1126.
31. Moyer, J.W., B.B. Cali, G.G. Kennedy and M.F. Abdou-Ghadir. 1980. Identification of two sweet

potato feathery mottle virus strains in North Carolina. *Plant Disease*, 64:762-764.

32. **Parrella, G., A. De Stradis and M. Giorgini.** 2006. sweet potato feathery mottle virus is the casual agent of sweet potato virus disease (SPVD) in Italy. *Plant Pathology*, 55: 818.
33. **Schaefer's, G.A. and E.R. Terry.** 1976. Insect transmission of sweet potato disease agents in Nigeria. *Phytopathology*, 66: 642-645.
34. **Souto, E.R., J. Sim, J. Chen, R.A. Valverde and C.A. Clark.** 2003. Properties of strains of sweet potato feathery mottle virus and tow newly recognized potyvirus infecting sweet potato in the United States. *Plant Disease*, 87: 1226-1232.
35. **Sulaiman, I., S. Zakaria and D. Yakoob.** 1998. Viral problems of sweet potato in Malaysia. *Agro-Search*, 5: 31- 34.
36. **Tairo, F., S.B. Mukasa, R.A.C. Jones, A. Kullaya, P.B. Rubaihayo and J.P.T. Valknen.** 2005. Unraveling the genetic diversity of the three main viruses involved in sweet potato viruses (SPVD), and its particle implications. *Molecular Plant Pathology*, 6: 199- 2110.
37. **Tugume, A.K., S.B. Mukasa and J.P.T. Valkonen.** 2008. Natural Wild Hosts of Sweet potato feathery mottle virus Show Spatial Differences in Virus Incidence and Virus-Like Diseases in Uganda. *Phytopathology*, 98: 640-652.
38. **Untiveros, M., S. Fuentes and J. Kreuze.** 2008. Molecular variability of sweet potato feathery mottle virus and other potyviruses infecting sweet potato in Peru. *Archives of Virology*, 156: 473- 483.
39. **Usugi, T., M. Nakano, O. Onuki, T. Maoka and T. Hayashi.** 1994. A new strain of sweet potato feathery mottle virus that causes russet crack on fleshy roots of some Japanese cultivars of sweet potato. *Annals of Phytopathology Society of Japan*, 60: 545- 554.
40. **Verdcourt, B.** 1963. *Convolvulaceae*. Pages 1-161 in *Flora of Tropical East Africa*. C.E. Hubbard and E. Milne-Redhead (eds.). Whitefriars Press Ltd., London.

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