

The Potency of Six Medicinal Plant Extracts Against the Stored Grain Insect Pest *Sitophilus granarius* L.

Nilesh Baburao Jawalkar^{1*}, Sureshchandra Popat Zambare¹ and Mohamed Izzat Al Ghannoum²

(1) Department of Zoology, Near Soneri Mahal, University Campus, Dr. Babasaheb Ambedkar Marathwada University, Aurangabad (MS) - 431004, India; (2) School of Natural and Environmental Sciences, Ridley Building, Newcastle University, NE1 7RU, United Kingdom. *Email of corresponding author: nbjawalkar@gmail.com

Abstract

Jawalkar, N.B., S.P. Zambare and M.I. Al Ghannoum. 2021. The Potency of Six Medicinal Plant Extracts Against the Stored Grain Insect Pest *Sitophilus granarius* L. Arab Journal of Plant Protection, 39(4): 323-328. <https://doi.org/10.22268/AJPP-039.4.323328>

This study was conducted on six medicinal plants viz., *Vitex negundo* (leaves), *Xanthium strumarium*, *Caesalpinia bonduc*, *Mucuna pruriens*, *Moringa oleifera* (seed kernels), *Tagetes erecta* (petals) for their bio-insecticidal activity. The powders of various parts of plants were extracted using the MARS6 microwave acid digestion system. Three different concentrations (20, 30, and 40%) of plant extracts were tested against granary weevil, *Sitophilus granarius* L. (Coleoptera: Curculionidae) for their bio-insecticidal activities under laboratory conditions. The results of statistical analysis showed a good performance of all plant extracts, especially at the high concentrations of the extracts, where they showed different levels of insect mortality and their developmental rate was also reduced leading to significant reduction in insect numbers. The mortality rate ranged was 0-100% with *S. granarius* adults ($p < 0.01$). There was significant correlation ($R=1$) between mortality rate and the concentration of plant extracts. However, the extract of *Mucuna pruriens* with solvents (Acetone + Petroleum Ether) showed the highest mortality rate of 100% at the three concentrations used ($p < 0.01$), whereas, the lowest average mortality of 43.3% ($p < 0.01$) was observed with *Xanthium strumarium* and *Vitex negundo* extracts with solvents (Methanol + n-Hexane) compared with 0% mortality in the control. All plant extracts have revealed insecticidal as well as propitious protective effect on grains, and it can be selected as effective control treatment after proper dose formulation to prevent weevil infestation in stored grains.

Keywords: Bioinsecticide, stored grains, *Sitophilus granarius*, mortality, medicinal plants, MARS 6.

Introduction

Cereal crops are affected by a wide range of factors during its different stages from the field up to the consumer, such as moisture, heat, insects, rodents and mite, which negatively change the characteristics of the physical and technological properties of the seeds with measurable losses in either quantity or quality (Kiaya, 2014). Globally, the estimated crop yield losses due to the crop insect pests were around 20-40%. Similarly, the reported world economic losses due to the different plant diseases is around 220 billion US Dollars per year (Agrios, 2005). Bradshaw *et al.* (2016) also reported the annual loss due to the invasive insect about 70 billion US dollars.

Stored grains are infested with insect pests causing approximately 10-25% losses worldwide (Raja *et al.*, 2001), poor seeds germination (Santos *et al.*, 1990), and reduction in their market value (Hill, 1990). In the tropics, Weaver & Petroff (2004) reported 30%, with losses net value in USA over 200 million US Dollars, annually.

FAO (1998) reported around 13×10^6 tons annual losses of stored grains caused by insects. Most of these insects are coleopteran species that thrive primarily on mould (Subramanyam & Harein, 1989; Viñuela *et al.*, 1993). The granary weevil, *Sitophilus granarius* L. is cosmopolitan in distribution. Both, adults and larvae cause damage to the stored grain (Fava & Burlando, 1995; Hill, 1990).

At present, the methods for controlling these pests are mainly based on the use of chemical insecticidal fumigants. The extensive use of synthetic insecticides for several decades led to long-term human health and environmental concerns, mainly because of their slow degradation rate in the environment and toxic residues, and the evolution of resistance to insecticides in insect pest populations (Isman, 2006) and consumer desire for pesticide-free grain (Arthur, 1996). According to estimates by the World Health Organization (WHO), there are around 200,000 deaths every year as a direct effect of poisoning with pesticides. Furthermore, the use of artificial chemicals nowadays became more restricted because of their high and acute toxicity, ability to create hormonal imbalance, spermatotoxicity, long degradation period, carcinogenicity, and accumulation as residues in food (Dubey *et al.*, 2011; Feng & Zheng, 2007; Khater, 2012a).

Universally, the environment is facing critical threat from the massive use of chemical pesticides. Therefore, it was very important to search for safer methods to manage insect pests which affect crops and stored grains (Khanzada *et al.*, 2015; Sarwar & Sattar, 2012). Hence, it is important to find suitable safe, effective, biodegradable, eco-friendly and low-cost alternatives to control the stored grain insect pests (Isman, 2006, 2014; Gandhi *et al.*, 2010), and many of such alternatives are now available (Dales, 1996; Defagó *et al.*, 2009; Isman, 2006). Identification of effective biodegradable botanical bio-products which do not harm

beneficial insects are now in progress (Haseeb *et al.*, 2004). There are many plants reported to possess pesticidal properties. These plants are a rich source of secondary metabolites, which might act on the insect physiological system and eventually control them (Daoubi *et al.*, 2005; Kim *et al.*, 2005). Locally available plants are currently in wide use to protect stored product against damage caused by insect infestation in many parts of the globe (Khater, 2012b). Monoterpenes and phenylpropenes found in these plants showed pesticidal activities have been tested for their larvicidal and bio-insecticidal property against several pests of stored products (Abdelgaleil *et al.*, 2009; Grodnitzky & Coats 2002; Lee *et al.*, 2003; Rice & Coats 2003; Cárdenas-Ortega *et al.*, 2005; Wuryatmo *et al.*, 2003; Duke *et al.*, 2000; Singh *et al.*, 2002; Lo Cantore *et al.*, 2009; Cristani *et al.*, 2007). The aim of the present investigation was to evaluate the bio-insecticidal effect of six Indian plant extracts classified as containing secondary bioactive compounds, extracted through MARS-6 instrument against the adults of *S. granarius*.

Materials and Methods

S. granarius rearing conditions

Adult insects of *S. granarius* were reared on wheat grains under laboratory conditions of $25\pm 1^\circ\text{C}$ temperature and relative humidity of $64\pm 5\%$, with L:D (12 h:12 h) photoperiod in glass cylindrical containers closed by metallic net. The insects were provided by the Food and Environment Research Agency (FERA), Fera Science Ltd., National Agri-Food Innovation Campus and Hutton, York, YO411LZ, United Kingdom. Adult weevils obtained from these cultures were used in the bioassay experiment.

Plant Materials

Fine powder of six samples from different parts of plants brought from the rural area near Aurangabad city, Maharashtra state, India for use in this experiment. Leaves, petals, and seed kernels were air dried naturally, then crushed into small pieces and then pulverized into fine powder. These plants were: *Vitex negundo* (leaves), *Xanthium strumarium*, *Caesalpinia bonduc*, *Mucuna pruriens*, *Moringa oleifera* (kernels), and *Tagetes erecta* (petals).

Preparation of extracts using MARS 6 System

The microwave system MARS 6 based on adding two solvents at once to samples to obtain the extracts was used. 0.5 g of each powder was added to two solvents (10 ml of Methanol, and 10 ml of Hexane) in a glass tube and placed in system for 40 minutes. The same procedure and the same amount were used again with another two solvents, acetone and petroleum ether. The excess solvents were evaporated during 48 hrs. and the extract was filtered to remove residual plant materials. The extract was then diluted with distilled water to obtain different concentrations (20, 30, and 40%) used in the experiment (Al-Ghannoum & Karso, 2015; Singh, 1994).

Bioassay, experimental design and statistical analysis

Completely randomized design (CRD) was used in this experiment. Three concentrations (20, 30, and 40%) were prepared from extracts. Filter papers in petri dishes were treated with 10 ml of each extract. The control was treated with water only. Ten adults of granary weevil, *S. granarius* were brought from stock culture and placed in petri dishes (9 cm diameter) containing 5 g wheat grains and 5 g wheat flour. Adults were left at room temperature (24°C) and 65-70% RH. All these cultures were used to evaluate the efficacy of these plant extracts, and dead insects of *S. granarius* in each petri dish were noted and discarded. The results of adult mortality rate were determined 72 hours after exposure.

Results and Discussion

The differences among the plant extracts, extract concentrations, and solvents, tested were determined using analysis of variance test (One-way ANOVA). The data obtained showed significant variation ($P < 0.01$). There were highly significant differences in the influence of plants on the mortality rate of *S. granarius* adults ranged from 0 to 100% ($P < 0.01$), with an average of 43.3-100% ($P < 0.01$). The results obtained showed a significant difference due to different concentrations of the plant extract on adult's mortality rate (Figure 1). The mortality rate ranged from 0% for the low concentration of extracts (20%) and reached 100% for the high concentration of the extracts (40%) for all five plant extracts (Figure 2). The adult's mortality rate increased with increase in concentration. The results proved that the concentration 40% of all plant extracts was very effective in increasing adults mortality rate, which ranged from 70-100% (average 88.3%) with methanol + n-hexane and 90-100% mortality (average 96.7%) with the solvents acetone + petroleum ether (Figure 3).

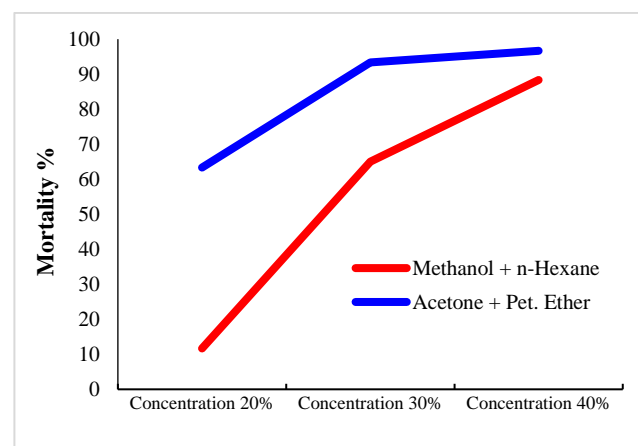


Figure 1. Average mortality rate *Sitophilus granarius* caused by three different concentration of plant extracts following the use of two groups of combined solvents. The first group included extracts obtained by using methanol and n-Hexane solvents, and in the second group, the extracts used were obtained by using acetone and petroleum ether solvents.

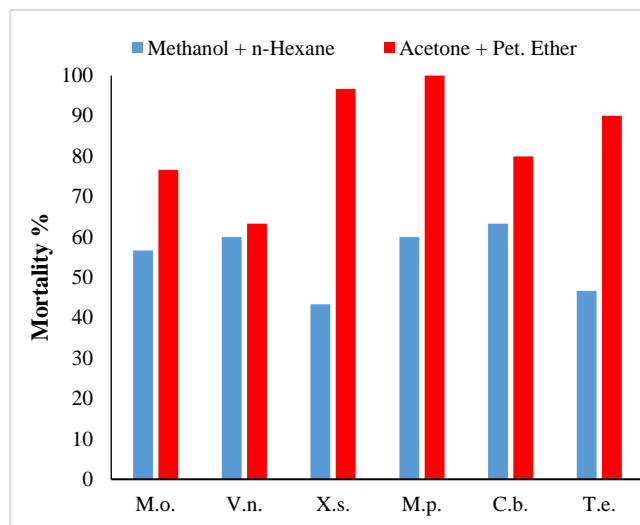


Figure 2. Average mortality rate of *Sitophilus granarius* following the use of six plant extracts by using two groups of combined solvents. The first group included extracts obtained by using methanol and n-hexane solvents, and in the second group, the extracts used were obtained by using acetone and petroleum ether solvents. Both groups showed their individual potential efficacy against the insect. Plants used were: M.o. (*Moringa oleifera*); V.n. (*Vitex negundo*); X.s. (*Xanthium strumarium*); M.p. (*Mucuna pruriens*); C.b. (*Caesalpinia bonduc*); T.e. (*Tagetes erecta*).

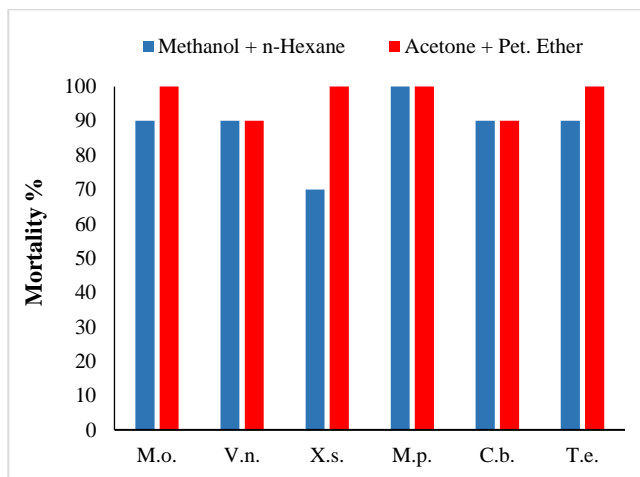


Figure 3. Mortality rate of *Sitophilus granarius* caused by using six plant extracts and using two groups of solvents (methanol + n-hexane and acetone + petroleum ether solvents) at extracts concentration of 40%. Plants used were: M.o. (*Moringa oleifera*); V.n. (*Vitex negundo*); X.s. (*Xanthium strumarium*); M.p. (*Mucuna pruriens*); C.b. (*Caesalpinia bonduc*); T.e. (*Tagetes erecta*).

The results reported here are in conformity with an earlier report which evaluated the effect of different concentrations of *Nerium oleander* leaves extracts on *Tribolium castaneum* adults (Alghannoum & Karso, 2015). In the present study, significant difference was observed on the mortality rate when extraction was made by different

solvents. The extracts obtained by the acetone + petroleum ether showed superiority in causing insect adults mortality rate of 63.3, 93.3, and 96.7% at 20, 30 and 40% extract concentrations, respectively (Figure 4), compared with 11.7, 65 and 96.7% mortality for using the solvents methanol + n-hexane with the same extracts concentrations, respectively (Figure 5). The minimum mortality rate of 0, 60 and 70% was observed with the same three plant extract *Xanthium strumarium* concentrations and using the solvents methanol + n-hexane, respectively. Whereas the highest mortality rate was obtained for the *Mucuna pruriens* plant extract and using acetone + petroleum ether solvents which led to 100% mortality for all three extract concentrations. These results are compatible with the results obtained earlier on the potential of *Mucuna pruriens* extract for its insecticidal activity against the adult beetles *Tribolium castaneum* (Hbst.) and cytotoxic activity against *Artemia salina* L. (Chhabi *et al.*, 2017).

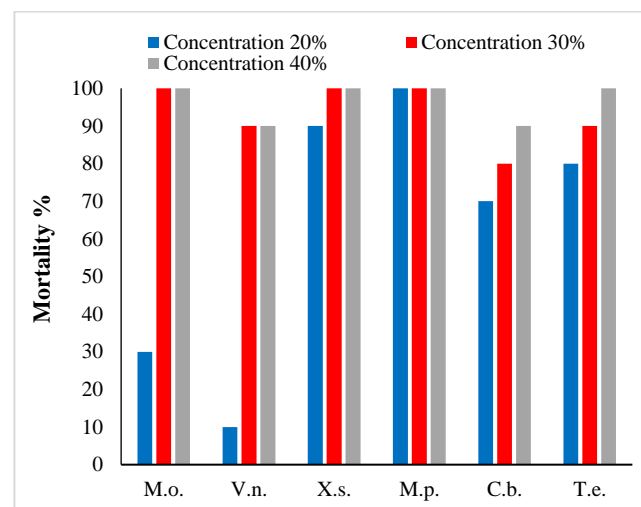


Figure 4. Average mortality rate of *Sitophilus granarius* following the use of six plant extracts with combined two solvents acetone + petroleum ether at 20%, 30% and 40% concentration. Plants used: M.o. (*Moringa oleifera*); V.n. (*Vitex negundo*); X.s. (*Xanthium strumarium*); M.p. (*Mucuna pruriens*); C.b. (*Caesalpinia bonduc*); T.e. (*Tagetes erecta*).

In addition to many medical benefits of *Mucuna pruriens* mentioned earlier (Lampariello, *et al.*, 2012; Yadav *et al.*, 2017), the plant contains a wide range of phytochemical constituents such as tannins, flavonoids, alkaloids and phenolic compounds which are responsible for different physiological effect and pharmacological activities (Deokar *et al.*, 2016). In the current investigation, mortality rate was significantly demonstrated for all six experimental plants extracts their efficiency against *Sitophilus granarius* insect pest. In many countries, the mixing of plant parts with grains is an ancient practice to manage stored grain insect pests (Kiruba *et al.*, 2008; Paul *et al.*, 2009). In such cases the larvae avoided feeding, and were not able to bore inside the grains, which can be due to the secondary metabolites present in the plants.

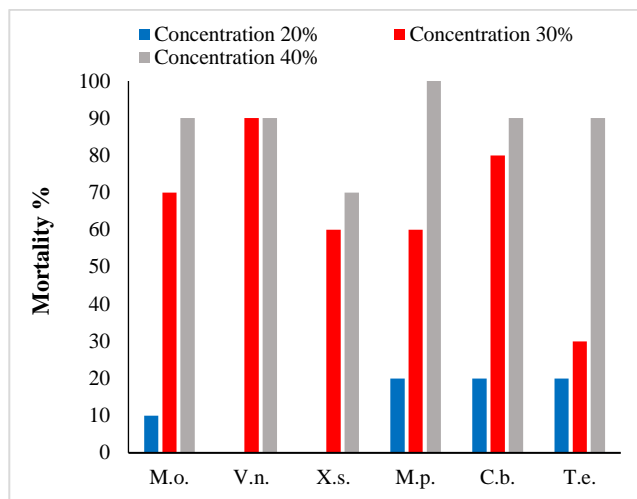


Figure 5. Average mortality rate of *Sitophilus granarius* following the use of six plant extracts and using methanol + n-hexane solvents at three different extract concentrations (20%, 30% and 40%). Plants used were: M.o. (*Moringa oleifera*); V.n. (*Vitex negundo*); X.s. (*Xanthium strumarium*); M.p. (*Mucuna pruriens*); C.b. (*Caesalpinia bonduc*); T.e. (*Tagetes erecta*).

In particular, the current six plants extracts were highly effective against insects, and contained compounds with insecticidal properties (Oelrichs *et al.*, 1983). Globally,

herbal insecticides are attracting more attention as they are considered suitable and good alternative to chemical pesticides. Therefore, naturally occurring environmental friendly biopesticide could be an alternative for chemical pesticides. In support of our previous study, the extracts of six plants in this study have shown bio-pesticidal and grain protective effects along with reduced growth of *S. granarius* adults suggesting bio-efficacy of these six medicinal plant extracts to manage post-harvest grain losses during long term storage. Furthermore, the microwave digestion system MARS 6 was capable in providing fast, complete digestion for lower detection limits in a high-throughput environment as compared to Soxhlet's apparatus.

The results of the present study revealed that all the extracts obtained through MARS 6 system have shown different levels of insecticidal potential against *S. granarius*. The 40% concentration of all plant extracts was very effective in inducing adult's mortality of the insect pest.

Acknowledgement

The authors wish to express their gratitude to the School of Natural and Environmental Sciences, Newcastle University for their laboratory facility. The financial assistance from Department of Science and Technology, Government of India and British Council, United Kingdom under Newton-Bhabha Fund are gratefully acknowledged.

المخلص

جاولكار، ن.ب.، س.ب. زامبارا وم.ع. الغنوم. 2021. فاعلية مستخلصات ستة نباتات طبية ضد آفة حشرة الحبوب المخزونة (*Sitophilus granarius* L.) مجلة وقاية النبات العربية، 39(4): 328-323. <https://doi.org/10.22268/AJPP-039.4.323328>

أجريت الدراسة على النباتات الطبية الستة التالية: أوراق *Vitex negundo*، بذور *Mucuna pruriens*، *Caesalpinia bonduc*، *Xanthium strumarium*، *Moringa oleifera*، وبتلات *Tagetes erecta*، وذلك لتقييم فعاليتها كمبيدات حيوية على حشرات الحبوب المخزونة. تم استخلاص المواد الفعالة من المساحيق الجافة لأجزاء مختلفة من النباتات باستخدام نظام الميكروويف MARS6 (تعمل درجات الحرارة المرتفعة والأوعية المغلقة والهضم الحمضي على تحضير العينات في وقت أقل من الطرائق التقليدية، وتستخدم حمضاً أقل، وتحتفظ حتى بالعناصر المتطايرة). تم اختبار ثلاثة تراكيز مختلفة من المستخلصات النباتية (20، 30 و 40%) ضد سوسة الحبوب (*Sitophilus granarius* L. (Coleoptera: Curculionidae)). أظهرت نتائج التحليل الإحصائي أداء جيداً لجميع المستخلصات النباتية، وبخاصة عند التراكيز العالية للمستخلصات، حيث أظهرت تفاوتاً في معدل نفوق الحشرات وانخفاض معدل نموها، مما أدى إلى انخفاض معنوي في أعداد الحشرات. تراوحت نسبة النفوق ما بين 0-100% عند البالغات ($p < 0.01$). أثبتت الدراسة وجود ارتباط معنوي ($R = 1$) وزيادة نسبة النفوق بشكل مباشر مع زيادة تركيز المستخلصات النباتية. ومع ذلك، أظهر مستخلص *M. pruriens* مع المذيبات Acetone + Petroleum Ether أعلى نسبة نفوق عند التركيزات الثلاثة 100% ($p < 0.01$)، بينما بلغ أقل معدل نفوق 43.3% ($p < 0.01$) لوحظ مع مستخلص *X. strumarium* و *V. negundo* مع المذيبات Methanol + n-Hexane مقارنة مع 0% عند الشاهد. أظهرت جميع المستخلصات النباتية قدرتها الوقائية كمبيدات كيميائية، ويمكن اعتمادها كخيار جيد بديل عن المبيدات الكيميائية عندما تحضر بالتراكيز المناسبة لمنع الإصابات الحشرية في الحبوب المخزونة.

كلمات مفتاحية: المبيدات الحيوية، الحبوب المخزونة، *Sitophilus granarius*، الوفيات، النباتات الطبية، MARS 6.

عناوين الباحثين: ن.ب. جاولكار^{1*}، س.ب. زامبارا¹ وم.ع. الغنوم². (1) قسم الحيوان، جامعة الدكتور بابا صاحب أمبيدكار ماراثوادا، الهند؛ (2) كلية العلوم الطبيعية والبيئية، جامعة نيوكاسل، المملكة المتحدة. *البريد الإلكتروني للباحث المرسل: nbjawalkar@gmail.com

References

- Abdelgaleil, S.A., M.I. Mohamed, M.E. Badawy and S.A. El-Arami.** 2009. Fumigant and contact toxicities of monoterpenes to *Sitophilus oryzae* (L.) and *Tribolium castaneum* (Herbst) and their inhibitory effects on acetylcholinesterase activity. *Journal of Chemical Ecology*, 35(5): 518-525.
<https://doi.org/10.1007/s10886-009-9635-3>
- Agrrios, G.N.** 2005. *Plant pathology* 5th Edition: Elsevier Academic Press. Burlington, Ma. USA: 79-103.
- Al-Ghannoum, M.I. and B.A. Karso.** 2015. Biological Potency of *Nerium oleander* L. Leaf Extracts on Mortality of the Red Flour Beetle *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). *Egyptian Journal of Biological Pest Control*, 25(1): 135.
- Arthur, F.H.** 1996. Grain protectants: current status and prospects for the future. *Journal of Stored Products Research*, 32(4): 293-302.
[https://doi.org/10.1016/S0022-474X\(96\)00033-1](https://doi.org/10.1016/S0022-474X(96)00033-1)
- Bradshaw, C.J., B. Leroy, C. Bellard, D. Roiz, C. Albert, A. Fournier, M. Barbet-Massin, J.M. Salles, F. Simard and F. Courchamp.** 2016. Massive yet grossly underestimated global costs of invasive insects. *Nature Communications*, 7(1): 1-8.
<https://doi.org/10.1038/ncomms12986>
- Cárdenas-Ortega, N.C., M.A. Zavala-Sánchez, J.R. Aguirre-Rivera, C. Pérez-González and S. Pérez-Gutiérrez.** 2005. Chemical composition and antifungal activity of essential oil of *Chrysactinia mexicana* Gray. *Journal of Agricultural and Food Chemistry*, 53(11): 4347-4349.
<https://doi.org/10.1021/jf040372h>
- Chhabi, S.B., M. Abdullah, K. Hasan, K. Tasnova, M. Khatun and N. Islam.** 2017. Potentiation of *Mucuna pruriens* (L.) DC. for insecticidal activity, insect repellency and brine shrimp lethality tests under the laboratory conditions. *Journal of Entomology and Zoology Studies*, 5(6): 692-696.
- Cristani, M., M. D'Arrigo, G. Mandalari, F. Castelli, M.G. Sarpietro, D. Micieli, V. Venuti, G. Bisignano, A. Saija and D. Trombetta.** 2007. Interaction of four monoterpenes contained in essential oils with model membranes: implications for their antibacterial activity. *Journal of Agricultural and Food Chemistry*, 55(15): 6300-6308. <https://doi.org/10.1021/jf070094x>
- Dales, M.J.** 1996. A review of plant materials used for controlling insect pests of stored products. *NRI Bulletin No.* 65:84.
- Daoubi, M., A. Deligeorgopoulou, A.J. Macías-Sánchez, R. Hernández-Galán, P.B. Hitchcock, J.R. Hanson and I.G. Collado.** 2005. Antifungal activity and biotransformation of diisophorone by *Botrytis cinerea*. *Journal of Agricultural and Food Chemistry*, 53(15): 6035-6039. <https://doi.org/10.1021/jf050600n>
- Defagó, M.T., A. Mangeaud, V. Benesovsky, C. Trillo, C. Carpinella, S. Palacios and G. Valladares.** 2009. *Melia azedarach* extracts: A potential tool for insect pest management. *Recent Progress in Medicinal Plants*. Houston: Studium Press LLC. 17-33.
- Deokar, G., P. Deore and S. Kshirsagar.** 2016. Phytochemistry and pharmacological activity of *Mucuna pruriens*: A review. *Pharmaceutical and Biological Evaluations*, 3(1): 50-59
- Dubey, N.K., R.A. Shukla, A.S. Kumar, P.R. Singh and B.H. Prakash.** 2011. Global scenario on the application of natural products in integrated pest management programmes. *Natural Products in Plant Pest Management*, 1: 1-20.
- Duke, S.O., J.G. Romagni and F.E. Dayan.** 2000. Natural products as sources for new mechanisms of herbicidal action. *Crop Protection*, 19(8-10): 583-589.
[https://doi.org/10.1016/S0261-2194\(00\)00076-4](https://doi.org/10.1016/S0261-2194(00)00076-4)
- FAO (Food and Agriculture Organization).** 1998. Quarterly bulletin of statistics, (Q.B.S) 11(3/4) 13-33.
- Fava, A.T. and B.R. Burlando.** 1995. Influence of female age and grain availability on the ovipositional pattern of the wheat weevil *Sitophilus granarius* (Coleoptera: Curculionidae). *European Journal of Entomology*, 92: 421-421.
- Feng, W. and X. Zheng.** 2007. Essential oils to control *Alternaria alternata* in vitro and in vivo. *Food Control*, 18(9): 1126-1130.
<https://doi.org/10.1016/j.foodcont.2006.05.017>
- Gandhi, N., S. Pillai and P. Patel.** 2010. Efficacy of pulverized leaves of *Punica granatum* (Lythraceae) and *Murraya koenigii* (Rutaceae) against stored grain pest, *Tribolium castaneum* (Herbst.) (Coleoptera: Tenebrionidae). *International Journal of Agriculture and Biology*, 12: 616-620.
- Grodniczky, J.A. and J.R. Coats.** 2002. QSAR evaluation of monoterpenoids' insecticidal activity. *Journal of Agricultural and Food Chemistry*, 50(16): 4576-4580.
<https://doi.org/10.1021/jf0201475>
- Haseeb, M., T.X. Liu and W.A. Jones.** 2004. Effects of selected insecticides on *Cotesia plutellae*, endoparasitoid of *Plutella xylostella*. *BioControl*, 49(1): 33-46.
<https://doi.org/10.1023/B:BICO.0000009377.75941.d7>
- Hill, D.S.** 1990. *Pests of stored products and their control*. Belhaven Press, London. 8-55.
- Isman, M.B.** 2006. Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Annual Review of Entomology*, 51: 45-66.
<https://doi.org/10.1146/annurev.ento.51.110104.151146>
- Isman, M.B.** 2014. Botanical insecticides: a global perspective. In *Biopesticides: State of the art and future Opportunities* American Chemical Society, 21-30.
<https://doi.org/10.1021/bk-2014-1172.ch002>
- Khanzada, H., M. Sarwar and M. Lohar.** 2015. Repellence activity of plant oils against red flour beetle *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae) in wheat. *International Journal of Animal Biology*, 1:86-92.

- Khater, H.F.** 2012a. Ecosmart biorational insecticides: alternative insect control strategies. *Advances in Integrated Pest Management*, 17-60.
- Khater, H.F.** 2012b. Prospects of botanical biopesticides in insect pest management. *Pharmacologia*, 3(12):641-656.
<https://doi.org/10.5567/pharmacologia.2012.641.656>
- Kiaya, V.** 2014. Post-harvest losses and strategies to reduce them. Technical Paper on Postharvest Losses. Action Contre la Faim (ACF), 25.
- Kim, H.G., J.H. Jeon, M.K. Kim and H.S. Lee.** 2005. Pharmacological effects of asaronaldehyde isolated from *Acorus gramineus* rhizome. *Food Science and Biotechnology*, 14(5):685-688.
- Kiruba, S., S. Jeeva, M. Kanagappan, S.I. Stalin and S.S. Das.** 2008. Ethnic storage strategies adopted by farmers of Tirunelveli district of Tamil Nadu, Southern Peninsular India. *Journal of Agricultural Technology*, 4(1): 1-10.
- Lampariello, L.R., A. Cortelazzo, R. Guerranti, C. Sticozzi and G. Valacchi.** 2012. The magic velvet bean of *Mucuna pruriens*. *Journal of Traditional and Complementary Medicine*, 2(4): 331-339.
[https://doi.org/10.1016/S2225-4110\(16\)30119-5](https://doi.org/10.1016/S2225-4110(16)30119-5)
- Lee, S., C.J. Peterson and J.R. Coats.** 2003. Fumigation toxicity of monoterpenoids to several stored product insects. *Journal of Stored Products Research*, 39(1): 77-85.
[https://doi.org/10.1016/S0022-474X\(02\)00020-6](https://doi.org/10.1016/S0022-474X(02)00020-6)
- Lo Cantore, P., V. Shanmugaiyah and N.S. Iacobellis.** 2009. Antibacterial activity of essential oil components and their potential use in seed disinfection. *Journal of agricultural and food chemistry*, 57(20): 9454-9461.
<https://doi.org/10.1021/jf902333g>
- Oelrichs, P.B., M.W. Hill, P.J. Vallely, J.K. MacLeod and T.F. Molinski.** 1983. Toxic tetranortriterpenes of the fruit of *Melia azedarach*. *Phytochemistry*, 22(2): 531-534.
[https://doi.org/10.1016/0031-9422\(83\)83039-8](https://doi.org/10.1016/0031-9422(83)83039-8)
- Paul, U.V., J.S. Lossini, P.J. Edwards and A. Hilbeck.** 2009. Effectiveness of products from four locally grown plants for the management of *Acanthoscelides obtectus* (Say) and *Zabrotes subfasciatus* (Boheman) (both Coleoptera: Bruchidae) in stored beans under laboratory and farm conditions in Northern Tanzania. *Journal of Stored Products Research*, 45(2): 97-107.
<https://doi.org/10.1016/j.jspr.2008.09.006>
- Raja, N., S. Albert, S.E. Ignacimuthu and S. Dorn.** 2001. Effect of plant volatile oils in protecting stored cowpea *Vigna unguiculata* (L.) Walpers against *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae) infestation. *Journal of Stored Products Research*, 37(2): 127-132.
[https://doi.org/10.1016/S0022-474X\(00\)00014-X](https://doi.org/10.1016/S0022-474X(00)00014-X)
- Rice, P.J. and J.R. Coats.** 2003. Insecticidal properties of several of several monoterpenoids to the house fly (Diptera: Muscidae), red flour beetle (Coleoptera: Tenebrionidae), and southern maize rootworm (Coleoptera: Chrysomelidae). *Journal of Economic Entomology*, 87: 1172-1179.
- Santos, J.P., J.D. Maia and I. Cruz.** 1990. Damage to germination of seed corn caused by maize weevil (*Sitophilus zeamais*) and Angoumois grain moth (*Sitotroga cerealella*). *Pesquisa Agropecuaria Brasileira*, 25(12):1687-1692.
- Sarwar, M. and M. Sattar.** 2012. Appraisal of Different Plant Products against *Trogoderma granarium* Everts to Protect Stored Wheat-A Laboratory Comparison. *The Nucleus*, 49(1): 65-69.
- Singh, H.P., D.R. Batish and R.K. Kohli.** 2002. Allelopathic effect of two volatile monoterpenes against bill goat weed (*Ageratum conyzoides* L.). *Crop Protection*, 21(4): 347-350.
[https://doi.org/10.1016/S0261-2194\(01\)00096-5](https://doi.org/10.1016/S0261-2194(01)00096-5)
- Singh, K.** 1994. Bioactivity of *Melia azedarach* L. against diamondback moth, *Plutella xylostella* L. (Doctoral dissertation, MSc thesis). Punjab Agricultural University, Ludhiana, India.
- Subramanyam, B.H. and P.K. Harein.** 1989. Insects infesting barley stored on farms in Minnesota. *Journal of Economic Entomology*, 82(6): 1817-1824.
<https://doi.org/10.1093/jee/82.6.1817>
- Viñuela, E., A. Adan, P. Del Estal, V. Marco and F. Budia.** 1993. Plagas de los productos almacenados. Hojas divulgadoras-Ministerio de Agricultura, Pesca y Alimentación, 1: 1-32.
- Weaver, D.K. and A.R. Petroff.** 2004. Pest management for grain storage and fumigation, Montana Department of Agriculture. Montana.
- Wuryatmo, E., A. Klieber and E.S. Scott.** 2003. Inhibition of citrus postharvest pathogens by vapor of citral and related compounds in culture. *Journal of Agricultural and Food Chemistry*, 51(9): 2637-2640.
<https://doi.org/10.1021/jf0261831>
- Yadav, M.K., P. Upadhyay, S. Purohit, B.L. Pandey and H. Shah.** 2017. Phytochemistry and pharmacological activity of *Mucuna pruriens*: A review. *International Journal of Green Pharmacy*, 11(2): 69-73.
<https://doi.org/10.22377/ijgp.v11i02.916>

Received: June 23, 2021; Accepted: September 30, 2021

تاريخ الاستلام: 2021/6/23؛ تاريخ الموافقة على النشر: 2021/9/30