

Biological and Chemical Control of Melon Root Rot Disease: A Review

E.A. Slebi* and H.H. Al-Juboory

Department of Plant Protection, College of Agricultural Engineering Sciences, University of Baghdad, Iraq.

*Email address of the corresponding author: emad.ali2204m@coagri.uobaghdad.edu.iq

Abstract

Slebi, E.A. and H.H. Al-Juboory. 2024. Biological and Chemical Control of Melon Root Rot Disease: A Review. Arab Journal of Plant Protection, 42(4): 465-473. <https://doi.org/10.22268/AJPP-001271>

Melon is an important crop grown in different regions of the world, and similar to many other crops, it is exposed to many diseases, including root rot, which is caused by a number of pathogens such as *Rhizoctonia solani*, *Fusarium solani*, *Macrophomina phaseolina*, *Fusarium oxysporum* f. sp. *melonis* (Fom), which causes Fusarium wilt and *Monosporascus* spp. which causes *Monosporascus* root rot and vine decline. To reduce the use of chemical pesticides because of their negative impact on health and the environment, biological control factors and some safe chemical agents are being increasingly used to control this disease. Many studies have shown the effectiveness of *Trichoderma* spp. and *Bacillus subtilis*, and vermicompost in reducing the disease by inhibiting the fungal growth of these pathogens present in the soil and inducing resistance in the melon plant against the disease. In addition, the use of algae was found to be efficient in the control of the disease. The use of some chemical agents, such as potassium salt (K_2HPO_4) and tannic acid, had a role in inhibiting some pathogens in the soil. This article summarizes research conducted to test the role of biological factors (vermicompost, microorganisms, and algae) as well as some chemical agents (tannic acid and K_2HPO_4) in controlling melon root rot disease.

Keywords: Melon root rot, biological control, K_2HPO_4 , vermicompost, algae.

Introduction

Melon, *Cucumis melo* L. is a major economic crop in the Cucurbitaceae family. It is grown around the world (Rocha *et al.*, 2010), including Iraq (Al-Juboory *et al.*, 2018). Soil-borne pathogens such as *Fusarium solani*, *Verticillium dahlia*, *Macrophomina phaseolina*, and *Rhizoctonia solani* can cause root rot in this crop (Buzi *et al.*, 2002; Infantino *et al.*, 2004). Disease agents like these are commonly controlled by using chemical fungicides (Kasumbwe *et al.*, 2014). Chemical fungicides are easy to use with high effectiveness, but they cause environmental pollution as well as the accumulation of chemicals in the food chain (Barák, 2017; Hussain *et al.*, 2009).

Agricultural pests, including pathogens and arthropods, cause significant losses annually (Adhab & Alkuwaiti, 2022; Adhab *et al.*, 2021; Al-Ani *et al.*, 2011b; Khalaf *et al.*, 2023; Schoelz *et al.*, 2021). Many studies demonstrated the effective use of biological control of pathogens present in the soil as a good alternative to chemical control, because it is safer and more environmentally friendly (Al-Ani *et al.*, 2021; 2013). One of these biological control agents that is used against these pathogens and pests is vermicompost; it is produced from the bio-oxidation process of many organic materials (plant and animal waste and sewage waste) by using various types of earthworms and microorganisms; it is rich in vitamins, microorganisms, growth hormones, and nutrients. (Ali *et al.*, 2015; Mupambwa & Mkeni, 2018; Sharma & Garg, 2018; Sulaiman & Mohamad, 2020).

Additionally, there are chemicals, such as potassium salt (K_2HPO_4), that reduce fungal pathogens population in the soil that cause damping-off, root rot, and wilting in beet

plants (Abdel-Monaim *et al.*, 2015). Tannic acid also greatly slows down the growth of fungal hyphae and the germination of *Fusarium oxysporum* conidia when tested in the laboratory (Wu *et al.*, 2010). Furthermore, other biological factors such as algae were also investigated as control agents for soil-borne diseases (Hewedy *et al.*, 2000). The blue-green algae and eukaryotic algae produce antifungal substances (Kulik, 1995; Schlegel *et al.*, 1998) that can inhibit the growth of many fungal pathogens (Bonjouklian *et al.*, 1991; Kiviranta *et al.*, 2006). Moreover, it was found that algae extract in powder (Weed-Max) or liquid form (Oligo-Mix) had the ability to inhibit fungal pathogens transmitted through soil. Algae are considered a good biological factor, along with other biological factors, in reducing root rot. (Abdel-Kader & El-Mougy, 2013). All these methods were effective in controlling plant diseases, including a number of crops such as melon, by inducing host resistance (Smith-Becker *et al.*, 2003), and thus reducing the effects of pathogens in the soil (Okubara & Paulitz, 2005).

In conclusion, chemical pesticides are dangerous to health and the environment, so this review summarizes the importance of chemical and biological agents for the control of melon root rot disease as an alternative and safer approach than chemical pesticides.

Melon crop value and importance

The origin of melon has been reported to be from China in the past two thousand years BC (Keng, 1974). It is a member of the Cucurbitaceae family. It includes numerous commercially significant crops, including diverse horticultural and botanical types that are widely cultivated across subtropical, tropical, and temperate regions of the world (Napolitano *et al.*, 2020). There are many uses for melon, as food, juice, nuts (seeds), and for medical use

(Robinson & Decker-Walters, 1997). In terms of global production, China is in the lead, with its production reaching 17 million tons, followed by the American continent, with its production reached 3.6 million tons (FAO, 2019). Worldwide, melon is cultivated in an area estimated at around 1.1 million hectares, with annual production reaching about 28 million tons (FAO, 2020). In Iraq, melon crops are planted on around 13.77 thousand hectares, with a yield of around 12.547 tons per hectare (Arab Organization for Agricultural Development, 2020).

Melon fruit is rich in carbohydrates, protein, vitamins, and minerals such as potassium (Shafeek *et al.*, 2015). In addition, melon contains many antioxidant compounds. These compounds have the ability to protect the cells of the human body against cancer with low calories and fat content (about 17 kcal/100 g) (Shafeek *et al.*, 2015). Among the most potent antioxidants are ascorbic acid, carotenoids, and phenolic compounds. Seeds contain vitamin E and omega-3 fatty acids (Menon & Rao, 2012). Melons also contain alkaloids, and flavonoids, which increase their health benefits for people (Chang *et al.*, 2015; Gómez-García *et al.*, 2020).

Melon diseases

Melon, like many other crops, is susceptible to several diseases. One of these diseases that reduces melon yields globally is the fungus *Fusarium oxysporum* f. sp. *melonis* (Fom), which causes the Fusarium wilt disease (Silvia *et al.*, 2017). Moreover, one of the main genera of fungi in the soil that cause root and seed rot disease is *Macrophomina* sp. (Botryosphaeriaceae), which affects more than 800 host plants worldwide (Paiva *et al.*, 2022).

One of the fungi that most often causes melon, *Cucumis melo* root rot or discolored vascular tissue is *Fusarium solani* (Aegerter *et al.*, 2000). Another disease is *Monosporascus* root rot and vine decline (MRRVD) caused by *Monosporascus* spp., which is currently one of the main obstacles to the production and spread of watermelon and melon crops globally (Bruton, 1998; Cohen *et al.*, 2012a; Martyn & Miller, 1996). The major symptoms of MRRVD on cucurbits appear just before harvest, when the leaves start to show yellowing, wilting and dryness followed by a dramatic drop in vine health and death, which results in a large economic loss (Cohen *et al.*, 2012a).

Among the common and destructive diseases that attack cucurbit leaves is powdery mildew (McGrath, 2017). The two most significant cucurbit powdery mildew (CPM) pathogen species are *Golovinomyces orontii* and *Podosphaera xanthii* (Kuzuya *et al.*, 2006). Mycelia and conidia are white in color, which mostly appear on stems and leaves but can also impact floral structures and fruits, as signs of infection. Leaf tissue that has been severely diseased may turn necrotic or chlorotic and brittle, and thus lowers photosynthetic capacity and consequently reduces yield and fruit quality (Stadnik & Bettiol, 2001). Another serious disease is downy mildew caused by the oomycete fungus *Pseudoperonospora cubensis* (Savory *et al.*, 2011).

Many nematodes affect vegetable crops, but the root Knot nematode *Meloidogyne* spp. is perhaps the most serious and severely attacks cucumber and melon (Bertrand, 2001).

Phytopathogen *Acidovorax citrulli* is the causal agent of the economically important disease known as bacterial fruit blotch (BFB), which affects the production of cucurbits. *A. citrulli* that infects the seed coat and spreads infection to the germinating seedling (Burdman & Walcott, 2012). BFB can appear in melons and other cucurbits at any growth stage. Water-soaked lesions on hypocotyls, cotyledons, and leaves are among the signs of BFB infection. Little and erratic water-soaked lesions on fruits spread through the peel, causing decays, fissures, and total product loss (Cunty *et al.*, 2019).

Melon root rot disease

Root rot diseases remain a major global threat to agricultural crop productivity. They are usually caused by more than one type of pathogen and are therefore often referred to as the root rot complex. Fungal and especially oomycete species are considered the main causes of the formation of this disease complex, and bacteria and viruses are also among the causes of root rot (Williamson-Benavides & Dhingra, 2021). Melon, like any other crop, is susceptible to infection with a variety of pathogens, and among these are soil-borne pathogens such as *Fusarium solani*, *Verticillium dahlia*, *Macrophomina phaseolina*, and *Rhizoctonia solani* that cause root rot (Buzi *et al.*, 2002; Hashem *et al.*, 2017; Infantino *et al.*, 2004; Jabr & Al-Jubouri, 2014; Matloob & al-Amiri, 2017). The fungi *M. cannonballus* and *M. eutypoides* are two of the most important causes of root rot and vine decline in melons worldwide, and *Fusarium oxysporum* f. sp. *melonis* vascular wilt disease affects the world's melon production (Castro *et al.*, 2020; Najeem & Kareem, 2018; Silvia *et al.*, 2017).

Failure to follow appropriate crop rotation enhances the survival of pathogens in the soil, which leads to a high infection rate and a decrease in crop yield and quality, as a result of the root system damage, which leads to the deterioration of the plant and consequently its death. In addition, these fungi remain in the soil or crop residue for long periods of time, they form reproductive structures and stone bodies that are resistant to unfavorable environmental conditions (Oumouloud *et al.*, 2013).

In response to the failure to deliver nutrients through vascular bundles because of root rot induced by the fungal pathogen *Monosporascus* sp. and causes yellowing of leaves and wilting of the plants (Cluck *et al.*, 2009; Picó *et al.*, 2008). For the first time in Iraq, Najeem & Kareem (2018) identified *M. cannonballus* as the causal agent of melon root rot.

The fungus *Macrophomina phaseolina* is one of the causes of fungal diseases found in the soil, which causes charcoal rot disease, which affects more than 500 plant species, including melon *Cucumis melo*, which again leads to rotting of the roots. The occurrence and severity of the disease is affected by the genetic nature of the cultivated variety, the age of the plant, temperature, and water availability (Cohen *et al.*, 2022).

Root rot disease often affects the productivity of melon crops in Brazil, and several *Fusarium* species were found to be the cause of this disease. 31 *Fusarium* isolates similar in morphology to *Fusarium solani* were collected from melon plants that showed root rot symptoms. Genetic analysis of

these isolates showed that 29 isolates belong to the fungus *Fusarium falciforme* and 2 isolates belong to the fungus *F. suttonianum*. The pathogenicity test showed that all isolates cause watermelon root rot and there are no significant differences between these isolates in terms of virulence (Silva *et al.*, 2003).

Chemical control of root rot disease

Chemicals such as potassium salt (K_2HPO_4) are effective in reducing the incidence of diseases caused by soil-borne fungi, including damping-off disease, root rot, and beet wilt (Abdel-Monaim *et al.*, 2015; Aleandri *et al.*, 2010). Previous research showed that seeds treated with acybenzolar-S-methyl (BTH) and dipotassium hydrogen phosphate (K_2HPO_4) produced plants that were more resistant to the pathogen *M. cannonballus* than seeds treated with methyl jasmonate, and reduced root rot disease severity. These products enhanced resistance through the production of a number of disease-related proteins (PR) that play a role in the resistance of the root system to the disease caused by the fungus *M. cannonballus*. It is known now that there is a complex pathway inside the plant cell that induces the production of hormones and enzymes that can inhibit the pathogen growth (Adhab, 2021). Previously reported studies conducted in the field and under greenhouse conditions showed that treatment with K_2HPO_4 induced resistance in beet plants against pathogenic fungi that cause damping off, root rot, and wilt, namely *R. solani*, *F. solani*, and *F. oxysporium* (Abdel-Monaim *et al.*, 2015).

Tannic acid, one of the tannins and a member of the phenolic acids family, is another chemical compound that has ten molecules of gallic acid connected to its core glucose molecule (Aelenei *et al.*, 2009). In both laboratory and field experiments, Bien (2016) demonstrated that tannic acid inhibited the growth of the fungus *F. graminearum*, which causes Fusarium head blight (FHB) on barley. In addition, tannic acid was also shown by Osman *et al.* (2019) to have an inhibitory effect on the *R. solani* fungus in both the laboratory and in the field. In many crops, including melon, chemically induced resistance to plant diseases is an efficient effect on disease control (Okubara & Paulitz, 2005; Smith-Becker *et al.*, 2003).

Biological control as an alternative to fungicides

Chemical fungicides were used to control root rot diseases because they are effective and easy to use (Kasumbwe *et al.*, 2014). However, these chemicals are expensive, can induce the emergence of pesticide-resistant plant pathogenic strains and pollutes the environment (Seong *et al.*, 2017). Accordingly, alternative, environment-friendly solutions need to be developed. Research showed that the biological management of diseases found in the soil is an effective safe substitute for chemical control (Al-Ani *et al.*, 2011a; 2011c; 2013).

Using natural farming products, such as vermicompost and its variants made from earthworms' metabolic fluids, mucus, and skin secretions, and bacteria that break them down, makes the soil more fertile and reduces the biological stress that soil pathogens put on plants (Gudeta *et al.*, 2021). Vermicompost useful components such as phosphorus bacteria, gibberellins, cytokinins, auxins, and Rhizobium

bacteria are needed for plant's growth. Vermicompost is rich in the major element's phosphorus 1.55-2.25%, potassium 1.85-2.25%, and nitrogen 2-3%, as well as the microelements necessary for plants (Edwards *et al.*, 2004; Sinha *et al.*, 2010). Vermicompost has the potential to inhibit infection, which makes it a superior option to chemical pesticides; this is because vermicompost is connected with earthworm excretions, especially skin secretions that include the antifungal substance lumbricin-PG (Li *et al.*, 2011; You *et al.*, 2019).

In both laboratory and field experiments, vermicompost inhibited the growth of *R. solani*, the fungus that causes cucumber damping off (Ersahin *et al.*, 2009; You *et al.*, 2019). *In-vitro* studies showed that when coelomic fluid from the earthworm *Eudrilus eugeniae* was added to the media, it inhibited the growth of four fungi: *Verticillium dahliae*, *Aspergillus flavus*, *R. solani*, and *F. oxysporium* (Rajesh *et al.*, 2019).

Vermicompost and the bacteria it contains, such as *Streptomyces* sp., *Pseudomonas* sp., *Burkholderia* sp., and *Bacillus* sp., can produce compounds that can suppress plant diseases (Sulaiman & Mohamad, 2020). In 2014, Jabr & Al-Jubouri reported that root and stem rot disease in melon plants is caused by a number of different fungi, such as *Drechslera australiansis*, *M. phaseolina*, *F. solani*, *F. oxysporium*, *F. proliferatum*, *R. solani*, and *Plectosphaerella cucumerina*. They also reported that *Azotobacter chroococcum* and *Pseudomonas fluorescens* were effective at suppressing these pathogens.

Hashem *et al.* (2017) showed that the bacteria *Bacillus subtilis* and the fungus *Trichoderma viride* could reduce the effects and frequency of charcoal rot in melons caused by the fungus *Macrophomina phaseolina*. It was found that treatments with salicylic acid, *Trichoderma harzianum*, and *B. subtilis* made root rot disease in melon caused by the fungus *R. solani* less common and less severe (Matloob & Al-Amri, 2017). According to Ahmed (2022), *Trichoderma harzianum* is a highly effective inhibitor of the fungi *F. oxysporium* and *Macrophomina phaseolina*, which are among the causes of root rot disease in melon plants.

The fungus *Trichoderma viride* Pers. is one of the powerful biological control agent that inhibits many plant pathogens, as it possesses enzymes that can decompose the cell walls of pathogenic fungi; It is also a good competitor for food and space, has the ability to produce antibiotics, and also works to induce systemic resistance in the plant and stimulate its growth (Sood *et al.*, 2020).

The use of yeasts to control pathogens is an additional bio-control method due to their ability to inhibit the growth of many pathogens; they are also considered safe for humans and the environment (Shalaby & El Nady, 2008). Fungal species like *Pythium* sp., *Fusarium* sp., and *Rhizoctonia solani* are inhibited by the yeasts *Saccharomyces cerevisiae* and *Kluyveromyces lactis* in the soil (Porhanife, 2010).

Hussein *et al.* (2022) showed that *Chaetomium cupreum*, *Trichoderma viride*, and *Saccharomyces cerevisiae* are bio-control agents that could effectively inhibit the growth of the soil-borne pathogens *Fusarium oxysporium*, *F. solani*, *Rhizoctonia solani*, *Pythium aphanidermatum*, and *Macrophomina phaseolina*. Such treatment reduced the infection rate and severity of root rot

disease in tomatoes, cucumbers, and cowpeas, and improved seed germination rate.

Another natural agent that can be used against pathogens is algae. *Chlorella vulgaris* and *Spirulina platensis* were found to reduce the symptoms of tomato root rot caused by the fungus *R. solani* (Muhanna *et al.*, 2015). In the laboratory, algae inhibited the growth of the fungal mycelium of *R. solani*, *F. oxysporum*, and *F. solani*. In greenhouse application, algae made it much less likely for seedlings to be killed pre- or post-germination compared to the control treatment (El-Sayed *et al.*, 2015).

At different locations, studies showed that commercial algae products treated with microorganisms (biological factors) protected tomato, cucumber, pepper, and melon plants from attack by root rot better than other treatments, such as chemical pesticides, with no significant increase in yield. The commercial algae product Weed-Max (2 g/L) + *Bacillus subtilis* showed superiority in reducing the incidence of the disease in vegetable plants over the treatment of the commercial algae product Oligo-X (2 ml/L) + *Trichoderma harzianum*, or the fungicide treatment (Abdel-Kader & El-Mougy, 2013). The blue-green algae extract had a role in inhibiting the growth of *Fusarium oxysporum* f. sp. *lycopersici*, the causal agent of tomato wilt in the laboratory and the field (Kim & Kim, 2008).

Integrated management of melon root rot disease

Integrated Pest Management (IPM) is a more recent eco-friendly approach for pest control where several control components are integrated with minimal use of chemical pesticides (Al-Ani *et al.*, 2021). Large reduction in melon productivity is caused by fungal diseases, mostly due to damage of the root system which can lead to plant death. Fungi are often difficult to control because they leave behind stable structures that can survive for a long period in the soil or crop residue such as sclerotia (*Rhizoctonia* sp., *Sclerotium* sp., and *Macrophomina* sp.) and chlamydozoospores (*Fusarium* sp.). When available, host resistance is the most appropriate control measure, but pathogens can develop new strains that can break down host resistant genes (Adhab *et al.*, 2018; Mohammed *et al.*, 2021; Oumouloud *et al.*, 2013). Soil fumigation with methyl bromide proved to be effective in controlling many soil-borne pathogens (Noling & Becker, 1994). However, methyl bromide application was banned internationally because of the environmental hazards it caused, and consequently encouraged the adoption of integrated disease/pest management. The environmentally friendly methods of managing root diseases include the

utilization of resistant varieties (Ambrósio *et al.*, 2015; Dias *et al.*, 2004; Salari *et al.*, 2012;), modification of irrigation systems (Cohen *et al.*, 2002), soil solarization (Cohen *et al.*, 2012b; Gamliel *et al.*, 1996; Guimarães *et al.*, 2008; Stapleton 2000), resistant rootstocks for grafting onto melon (Cohen *et al.*, 2012b; Cohen *et al.*, 2002), biopesticides (Narayan *et al.*, 2015), and incorporation of green manure (Dantas *et al.*, 2013). All proved to have good control potential. Such approaches can better support a balanced soil microbiome and enhance disease management. Such approaches need to be investigated for the control of melon root rot. Plant residues can boost soil fertility and microbial biomass when they are added to the soil, and at the same time release chemicals that inhibit soil-borne pathogens (Stone *et al.*, 2004). In green manure farming, root pathogens can be controlled via biofumigation, in which poisonous compounds are generated when plant residue decomposes. Mulching has the ability to physically obstruct light in the soil, which is necessary for the development of pathogen structures, the emergence of dormant spores, and/or the development of resistant structures such as sclerotia (Lobo Junior *et al.*, 2009).

Conclusions

Melon is a major crop and is grown in many regions of the world and over large areas. It is infected with many fungal pathogens, which cause many diseases that affect yield and quality. Among these diseases is root rot disease, which is caused by many pathogens present in the soil, and belong to the genera *Fusarium*, *Rhizoctonia*, *Pythium*, *Macrophomina*, *Monosporascus*, and possibly others.

Chemical pesticides have played a role in controlling these diseases, including root rot, but for the negative effects of these pesticides on health and the environment and for the purpose of finding safe and environment-friendly alternatives in combating these diseases, some biological and physical factors have been successfully used (Al-Ani *et al.*, 2011b; 2011c). Among the biological factors are microorganisms such as *Trichoderma* sp., *Bacillus* sp. bacteria, and others. Vermicompost was successfully used to control the root rot disease by inhibiting the pathogens, improving plant growth and inducing host resistance. Furthermore, algae extracts were found to be an effective component for the control of the disease. The safe chemical agents found to effectively control the disease include tannic acid and potassium salt K_2HPO_4 .

المخلص

صليبي، عماد علي وحرية حسين الجبوري. 2024. المكافحة الحيوية والكيميائية لمرض تعفن جذور البطيخ: مراجعة علمية. مجلة وقاية النبات العربية، 42(4): 465-473. <https://doi.org/10.22268/AJPP-001271>

يعدّ البطيخ من المحاصيل المهمة والذي يزرع في مناطق مختلفة من العالم. يتعرض محصول البطيخ مثل باقي المحاصيل الأخرى إلى العديد من الأمراض، ومنها مرض تعفن الجذور الذي تسببه العديد من مسببات المرضية، مثل: *Fusarium solani*، *Rhizoctonia solani*، *Macrophomina phaseolina*، *Fusarium* *oxysporum* f. sp. *melonis* (Fom) الذي يسبب الذبول الفيوزاري و *Monosporascus* spp. الذي يسبب مرض تعفن الجذور وتدهور الأفرع. وللحدّ من استخدام المبيدات الكيماوية بسبب تأثيرها السلبي على الصحة والبيئة، يتم استخدام عوامل المكافحة الحيوية وبعض المركبات الكيميائية الآمنة على نحوٍ متزايد للسيطرة على

هذا المرض. أظهرت العديد من الدراسات فعالية كل من *Bacillus subtilis* و *Trichoderma* spp. والسماذ الدودي في الحد من المرض عن طريق تثبيط نمو الفطور الممرضة الموجودة في التربة وتحفيز مقاومة نبات البطيخ للمرض. بالإضافة إلى ذلك، وجد أن استخدام الطحالب ذو فعالية في السيطرة على المرض. وكان لاستخدام بعض المركبات الكيميائية، مثل ملح فوسفيت البوتاسيوم (K_2HPO_4) وحمض التانيك، دور في تثبيط بعض مسببات الأمراض في التربة. يلخص هذا المقال الأبحاث التي أجريت لاختبار دور العوامل الحيوية (السماذ الدودي، الكائنات الحية الدقيقة والطحالب) وكذلك بعض المركبات الكيميائية (حمض التانيك و K_2HPO_4) في السيطرة على مرض تعفن جذور البطيخ.

كلمات مفتاحية: تعفن جذور البطيخ، المكافحة الحيوية، K_2HPO_4 ، السماذ الدودي، الطحالب.

عناوين الباحثين: عماد علي صليبي* وحرية الجبوري، قسم وقاية النبات، كلية علوم الهندسة الزراعية، جامعة بغداد، العراق. *البريد الإلكتروني للباحث المراسل: emad.ali2204m@coagri.uobaghdad.edu.iq

References

- Abdel-Kader, M.M and N.S. El-Mougy.** 2013. Bioagents and commercial algae products as integrated biocide treatments for controlling root rot diseases of some vegetables under protected cultivation system. *Journal of Marine Biology*, 2013:429850. <https://doi.org/10.1155/2013/429850>
- Abdel-Monaim, M.F., M.A.M. Atwa and K.M. Morsy.** 2015. Induced systemic resistance against root rot and wilt diseases in fodder beet (*Beta vulgaris* L. var. *rapacea* Koch.) by using potassium salts. *Journal of Plant Pathology and Microbiology*, 6(10):315. <https://doi.org/10.4172/2157-7471.1000315>
- Adhab, M.** 2021. Be smart to survive: virus-host relationships in nature. *Journal of Microbiology, Biotechnology and Food Sciences*, 10(6):e3422. <https://doi.org/10.15414/jmbfs.3422>
- Adhab, M. and N.A. Alkuwaiti.** 2022. Geminiviruses occurrence in the Middle East and their impact on agriculture in Iraq. Pp. 171-185. *In: Geminivirus: Detection, Diagnosis and Management.* R.K. Gaur, P. Sharma and H. Czosnek (eds.). Elsevier, USA. <https://doi.org/10.1016/b978-0-323-90587-9.00021-3>
- Adhab, M., C. Angel, S. Leisner and J.E. Schoelz.** 2018. The P1 gene of Cauliflower mosaic virus is responsible for breaking resistance in Arabidopsis thaliana ecotype Enkheim (En-2). *Virology*, 523:15-21. <https://doi.org/10.1016/j.virol.2018.07.016>
- Adhab, M., N. Al-Kuwaiti and R. Al-Ani.** 2021. Biodiversity and occurrence of plant viruses over four decades: Case study for Iraq. Pp. 159-163. *In: Proceedings of the Third International Sustainability and Resilience Conference: Climate Change.* November 15-16, 2021, IEEE, Virtual Conference. <https://doi.org/10.1109/IEEECONF53624.2021>
- Aegerter, B.J., T.R. Gordon and R.M. Davis.** 2000. Occurrence and pathogenicity of fungi associated with melon root rot and vine decline in California. *Plant Disease*, 84(3):224-230. <https://doi.org/10.1094/pdis.2000.84.3.224>
- Aelenei, N., M.I. Popa, O. Novac, G. Lisa and L. Balaita.** 2009. Tannic acid incorporation in chitosan-based microparticles and in vitro controlled release. *Journal of Materials Science*, 20(5):1095-1102. <https://doi.org/10.1007/s10856-008-3675-z>
- Ahmed, A.M.** 2022. Evaluation of some biological and chemical control agents against *Fusarium oxysporum* and *Macrophomina phaseolina* that cause root rot disease on watermelon. M.Sc. thesis, College of Agricultural Engineering Sciences, University of Baghdad. 92 pp.
- AlAni, R., M.A. Adhab and S.A. Hamad.** 2011a. Evaluation of the efficiency of different techniques for extraction and purification of Tomato yellow leaf curl virus (TYLCV). *Baghdad Science Journal*, 8(1):447-452. <https://doi.org/10.21123/bsj.2011.8.1.447-452>
- Al-Ani, R., S.N. Diwan and M.A. Adhab.** 2011b. Systemic resistance induced in potato plants against Potato virus Y common strain (PVY) by plant extracts in Iraq. *Advances in Environmental Biology*, 5(2):209-215.
- Al-Ani, R.A., M.A Adhab, M.A. El-Muadhidi and M.A. Al-Fahad.** 2011c. Induced systemic resistance and promotion of wheat and barley plants growth by biotic and non-biotic agents against barley yellow dwarf virus. *African Journal of Biotechnology*, 10(56):12078-12084.
- Al-Ani, R.A., M.A. Athab and O.N. Matny.** 2013. Management of potato virus Y (PVY) in potato by some biocontrol agents under field conditions. *Journal of Pure and Applied Microbiology*, 7(4):2861-2865.
- Al-Ani, L.K.T., Surono, L. Aguilar-Marcelino, V. E. Salazar-Vidal, A. G. Becerra and W. Raza.** 2021. Role of useful fungi in agriculture sustainability. Pp. 1-44. *In: Recent Trends in Mycological Research.* A.N. Yadav (ed.), Springer, Cham. https://doi.org/10.1007/978-3-030-60659-6_1
- Aleandri, M.P., R. Reda, V. Tagliavento, P. Magro and G. Chilosi.** 2010. Effect of chemical resistance inducers on the control of *Monosporascus* root rot and vine decline of melon. *Phytopathologia Mediterranea*, 49(1):18-26. https://doi.org/10.14601/Phytopathol_Mediterr-3117
- Ali, U., N. Sajid, A. Khalid, L. Riaz, M.M. Rabbani, J.H. Syed and R.N. Malik.** 2015. A review on vermicomposting of organic wastes. *Environmental Progress and Sustainable Energy*, 34(4):1050-1062. <https://doi.org/10.1002/ep.12100>
- Al-Juboori, A.W.A., E.N. Ismail and K.A. Alwan.** 2018. Molecular and morphological indicators (Qutha) Cucumis melo planted in Iraq. *Iraqi Journal of Agricultural Sciences*, 50(3):835-841. <https://doi.org/10.36103/ijas.v50i3.700>
- Ambrósio, M.M.Q., A.C. Dantas, E. Martínez-Perez, A.C. Medeiros, G.H.S. Nunes and M.B. Picó.** 2015. Screening a variable germplasm collection of *Cucumis*

- melo* L. for seedling resistance to *Macrophomina phaseolina*. *Euphytica*, 206(2):287-300.
<https://doi.org/10.1007/s10681-015-1452-x>
- Arab Organization for Agricultural Development (AOAD)**. 2020. Yearbook of Agricultural Statistics, 41.
- Barák, I.** 2017. Editorial: Spores and spore formers, *Frontiers Microbiology*, 8:1046.
<https://doi.org/10.3389/fmicb.2017.01046>
- Bertrand, C.** 2001. Lutte contre les nématodes à galles (*Meloidogyne* spp.) en agriculture biologique. Groupe de Recherche en Agriculture Biologique (édition Janvier 2001):1-4.
- Bien, B.** 2016. A comparison of tannic acid biopesticide and A commercial fungicide used for crop protection against fusarium head blight. M.Sc. thesis, Chemistry Department, Western Carolina University, USA. 75 pp.
- Bonjouklian, R., T.A. Smitka, L.E. Doolin, R. Molloy, M. Debono, S.A. Shaffer, R.E. Moore, J.B. Stewart and G.M. Patterson.** 1991. Tjipanazoles, new antifungal agents from the blue-green alga *Tolypothrix tjipanansensis*, *Tetrahedron*, 47(37):7739-7750.
[https://doi.org/10.1016/S0040-4020\(01\)81932-3](https://doi.org/10.1016/S0040-4020(01)81932-3)
- Bruton, B.D.** 1998. Soilborne diseases in cucurbitaceae: Pathogen virulence and host resistance. Pp. 143-166. *In: Cucurbitaceae J. McCreight (ed.)*. International Society for Horticultural Science, Leuven, Belgium.
- Burdman S. and R. Walcott.** 2012. *Acidovorax citrulli*: Generating basic and applied knowledge to tackle a global threat to the cucurbit industry. *Molecular Plant Pathology*, 13(8):805–815.
<https://doi.org/10.1111/j.1364-3703.2012.00810.x>
- Buzi, A., G. Chilosi and P. Magro.** 2002. The main diseases of melon. *Culture Protette*, 31(9):31-45.
- Castro, G., G. Perpiñá, C. Esteras, J. Armengol, B. Picó and A. Pérez de Castro.** 2020. Resistance in melon to *Monosporascus cannonballus* and *M. eutypoides*: Fungal pathogens associated with *Monosporascus* root rot and vine decline. *Annals of Applied Biology*, 177(7):1-11. <https://doi.org/10.1111/aab.12590>
- Chang, C.I., C.H. Chou, M.H. Liao, T.M. Chen, C.H. Cheng, R. Anggriani, C.P. Tsai, H.I. Tseng and H.L. Cheng.** 2015. Bitter melon triterpenes work as insulin sensitizers and insulin substitutes in insulin-resistant cells. *Journal of Functional Food*, 13:214-224.
<https://doi.org/10.1016/j.jff.2014.12.050>
- Cluck, T.W., C.L. Biles, M. Duggan, T. Jackson, K. Carson, J. Armengol and B.D. Bruton.** 2009. Association of dsRNA to down-regulation of perithecial synthesis in *Monosporascus cannonballus*. *The Open Mycology Journal*, 3(1):9-19.
<https://doi.org/10.2174/1874437000903010009>
- Cohen, R., C. Horev, Y. Burger, S. Shirber, J. Hershenhorn, J. Katan and M. Edelstein.** 2002. Horticultural and pathological aspects of Fusarium wilt management using grafted melons. *HostScience*, 37(7):1069-1073.
- Cohen, R., M. Elkabetz, H. Paris, S. Freeman and A. Gur.** 2022. Charcoal rot (*Macrophomina phaseolina*) across melon diversity: evaluating the interaction between the pathogen, plant age and environmental conditions as a step towards breeding for resistance. *European Journal of Plant Pathology*, 163:601-613.
<https://doi.org/10.1007/s10658-022-02500-2>
- Cohen, R., N. Omari, A. Porat and M. Edelstein.** 2012b. Management of *Macrophomina* wilt in melons using grafting or fungicide soil application: pathological, horticultural and economical aspects. *Crop Protection*, 35:58-63.
<https://doi.org/10.1016/j.cropro.2011.12.015>
- Cohen, R., S. Pivonia, K.M. Crosby and R.D. Martyn.** 2012a. Advances in the biology and management of *Monosporascus* vine decline and wilt of melons and other cucurbits *Horticultural Reviews*, 39:77-120.
<https://doi.org/10.1002/9781118100592.ch2>
- Cunty, A., C. Audusseau, S. Paillard, V. Olivier, C. François, C. Rivoal and F. Poliakoff.** 2019. First report of *Acidovorax citrulli*, the causal agent of bacterial fruit blotch, on melon (*Cucumis melo*) in guadeloupe (France). *Plant Disease*, 103(5):1017-1017.
<https://doi.org/10.1094/PDIS-10-18-1825-PDN>
- Dantas, A.M.M., M.M.Q. Ambrósio, S.R.C. Nascimento, R.F. Senhor, M.A. Cézar and J.S.S. Lima.** 2013. Incorporation of plant materials in the control of root pathogens in muskmelon. *Revista Agro@mbiente On-line*, 7(3):338-344.
<https://doi.org/10.18227/1982-8470ragro.v7i3.1257>
- Dias, R.C.S., B. Picó, A. Espinos and F. Nuez.** 2004. Resistance to melon vine decline derived from *Cucumis melo* sp. *agrestis*: genetic analysis of root structure and root response. *Plant Breeding*, 123(1):66-72. <https://doi.org/10.1046/j.1439-0523.2003.00944.x>
- Edwards, C.A., J. Dominguez and N.Q. Arancon.** 2004. The influence of vermicomposts on plant growth and pest incidence. Pp. 397-420. *In: Soil Zoology for Sustainable Development in the 21st Century*. S.H. Shakir and W.Z.A. Mikhail (eds.). Cairo, Egypt.
- EL-Sayed, S.A. and A.M. Mousa.** 2015. Effect of some algal filtrates and chemical inducers on root-rot incidence of faba bean. *Agricultural Research and Technology*, 1(1):5-9.
<https://doi.org/10.19080/ARTOAJ.2015.01.555552>
- Ersahin, Y.S., K. Haktanir and Y. Yanar.** 2009. Vermicompost suppresses *Rhizoctonia solani* Kühn in cucumber seedlings. *Journal of Plant Diseases and Protection*, 116(4):182-188.
<https://doi.org/10.1007/BF03356308>
- FAOSTAT (Food and Agriculture Organization of the United Nations).** *Statistics Database*. 2019. <http://www.fao.org/faostat/en/#data>
- FAOSTAT (Food and Agriculture Organization of the United Nations).** *Statistics Database*. 2020. Retrieved from: <http://www.fao.org/faostat/en/#home>
- Gamliel, A., A. Grinstein and J. Katan.** 1996. Combining solarization and fumigants as feasible alternatives to methyl bromide. Pp. 17-18. *In: Proceedings of the Annual International Conference, Methyl Bromide Alternatives Emission Reduction*. November 3-6, 1996, Orlando Florida, USA.

- Gómez-García, R., D.A. Campos, C.N. Aguilar, A.R. Madureira and M. Pintado.** 2020. Valorization of melon fruit (*Cucumis melo* L.) by-products: Phytochemical and biofunctional properties with emphasis on recent trends and advances. *Trends in Food Science Technology*, 99:507–519. <https://doi.org/10.1016/j.tifs.2020.03.033>
- Gudeta, K., J.M. Julka, A. Kumar, A. Bhagat and A. Kumari.** 2021. Vermiwash: An agent of disease and pest control in soil: a review. *Heliyon*, 7(3):e06434. <https://doi.org/10.1016/j.heliyon.2021.e06434>
- Guimarães, I. M., R. Sales Junior, K. J. P. Silva, S. J. Michereff and D. R. S. Nogueira.** 2008. Efeito de fluazinam no controle ramas em meloeiro. *Revista Caatinga*, 21(4):147-153.
- Hashem, M.Z., S.H Samir and A.K. Hassan.** 2017. Detect activity of some biological factors to induce resistance in cantaloupe plant through peroxidase enzyme, phenols and chlorophyll contents. *Iraqi Journal of Agricultural Sciences*, 48(5):1239-1246. <https://doi.org/10.36103/ijas.v48i5.333>
- Hewedy, M.A., M.M.H. Rahhal and I.A. Ismail.** 2000. Pathological studies on soybean damping-off disease. *Egyptian Journal of Applied Sciences*, 15:88-102.
- Hussain, S.T., M. Siddique, M. Saleem, M. Arshad and A. Khalid.** 2009. Chapter 5 impact of pesticides on soil microbial diversity, enzymes, and biochemical reactions. *Advances in Agronomy*, 102:159-200. [https://doi.org/10.1016/S0065-2113\(09\)01005-0](https://doi.org/10.1016/S0065-2113(09)01005-0)
- Hussein, S.N., A.Z.J. Ali and H.H. Al-Juboory.** 2022. Evolution of some biological and chemical elements in controlling some soil borne fungi and stimulating plant growth. *Arab Journal of Plant Protection*, 40(1):37-47. <https://doi.org/10.22268/AJPP-40.1.037047>
- Infantino, A., A. Carlucci, N. Pucci, G. Ciuffreda, C. Montuschi, F. Lops, A. Uccelletti, M. Mucci and S. Frisullo.** 2004. Funghi che causano marciumi radicali e il collasso di Cucurbitaceae in Italia. *Petria*, 14:77-89.
- Jabr, K.S. and H.H. Al-Jubouri.** 2014. Evaluation of the efficiency of *Azotobacter chroococcum* and *Pseudomonas fluorescens* in controlling some watermelon root rot causes. *Egyptian Journal of Applied Sciences*, 19(5):113-131.
- Kasumbwe, K, K.N. Venugopala, V. Mohanlall and B. Odhav.** 2014. Antimicrobial and antioxidant activities of substituted halogenated coumarins. *Journal of Medicinal Plant Research*, 8(5):274-281. <https://doi.org/10.5897/JMPR2013.4419>
- Keng, H.** 1974. Economic plants of ancient north China as mentioned in Shih Ching (Book of Poetry). *Economic Botany*, 28:391-410. <https://doi.org/10.1007/BF02862856>
- Khalaf, L.K., M. Adhab, L.M. Aguirre-Rojas and A.E. Timm.** 2023. Occurrences of wheat curl mite *Aceria tosichella* keifer 1969 (eriophyidae) and the associated viruses (WSMV, HPWMOV, TriMV) in Iraq. *Iraqi Journal of Agriculture Sciences*, 54(3):837-849. <https://doi.org/10.36103/ijas.v54i3.1767>
- Kim, J. and J.D. Kim.** 2008. Inhibitory effect of algal extracts on mycelial growth of the tomato-wilt pathogen, *Fusarium oxysporum* f. sp. *lycopersici*. *Mycobiology*. 36(4):242-248. <https://doi.org/10.4489/MYCO.2008.36.4.242>
- Kiviranta, J., A. Abdel-Hameed, K. Sivonen, S.I. Niemelä and G. Carlberg.** 2006. Toxicity of cyanobacteria to mosquito larvae-screening of active compounds. *Environmental Toxicology and Water Quality*, 8(1):63-71. <https://doi.org/10.1002/tox.2530080107>
- Kulik, M.M.** 1995. The potential for using cyanobacteria (blue-green algae) and algae in the biological control of plant pathogenic bacteria and fungi. *European Journal of Plant Pathology*, 101(6):585-599. <https://doi.org/10.1007/BF01874863>
- Kuzuya, M., K. Yashiro, K. Tomita and H. Ezura.** 2006. Powdery mildew (*Podosphaera xanthii*) resistance in melon is categorized into two types based on inhibition of the infection processes. *Journal of Experimental Botany*, 57(9):2093-2100. <https://doi.org/10.1093/jxb/erj166>
- Li, W., S. Li, J. Zhong, Z. Zhu, J. Liu and W. Wang.** 2011. A novel antimicrobial peptide from skin secretions of the earthworm, *Pheretima guillelmi* (Michaelsen). *Peptides*, 32(6):1146–1150. <https://doi.org/10.1016/j.peptides.2011.04.015>
- Lobo Junior, M., R.S. Brandão, C.A. Corrêa, C.A. Görgen, E.A. Civardi and P.D. Oliveira.** 2009. Uso de braquiárias para o manejo de doenças causadas por patógenos habitantes do solo. *Embrapa Arroz e Feijão*. Comunicado Técnico, 8 pp.
- Martyn, R.D. and M.E. Miller.** 1996. *Monosporascus* root rot and vine decline: An emerging disease of melons worldwide. *Plant Disease*, 80:716-725. <https://doi.org/10.1094/PD-80-0716>
- Matloob, A.A.A.H. and E.Q.A. Al-Amiri.** 2017. Molecular diagnosis of *Rhizoctonia solani* Kühn caused of melon root *Cucumis melo* L. and its biological control. *Al Furat Journal of Agricultural Sciences*. 9(4):1121-1139.
- McGrath, M.T.** 2017. Powdery mildew. Pp 62-64. *In:* Compendium of Cucurbit Diseases and Insect Pests. Second edition. A.P. Keinath, W.M. Wintermantel and T.A. Zitter (eds.). The American Phytopathological Society Press. St. Paul, Minesota, USA. <https://doi.org/10.1094/9780890545744>
- Menon, S.V. and T. Rao.** 2012. Nutritional quality of muskmelon fruit as revealed by its biochemical properties during different rates of ripening. *International Food Research Journal*, 19(4):1621-1628.
- Mohammed, D., M. Adhab and N. Al-Kuwaiti.** 2021. Molecular characterization of viruses associated with leaf curl disease complex on zucchini squash in Iraq reveals Deng primer set could distinguish between new and old world Begomoviruses. *Anal. da Academia Brasileira de Ciências*, 93:e20210050. <https://doi.org/10.1590/0001-3765202120210050>
- Muhanna, N.A.S., S.S.M. Ragab and A.A. El Ghumm.** 2015. Biocontrol of tomato root-rot caused by *Rhizoctonia solani*. *Egypt Journal of Phytopathology*, 43(1):89-100. <https://doi.org/10.21608/ejp.2015.94578>

- Mupambwa, H.A. and P.N.S. Mnkeni.** 2018. Optimizing the vermicomposting of organic wastes amended with inorganic materials for production of nutrient-rich organic fertilizers: a review. *Environmental Science and Pollution Research*, 25(11):10577-10595. <https://doi.org/10.1007/s11356-018-1328-4>
- Najeem, H.W. and T.A. Kareem.** 2018. Morphological and molecular identification of *Monosporascus cannonballus* causal agent of melon root rot and plant decline in Iraq. *Journal of Biodiversity and Environmental Science*, 13(6):83-88.
- Napolitano, M., N. Terzaroli, S. Kashyap, L. Russi, E. Jones-Evans and E. Albertini.** 2020. Exploring Heterosis in Melon (*Cucumis melo* L.). *Plants*, 9(2):282. <https://doi.org/10.3390/plants9020282>
- Narayan, S., V. Kumar and S. Singh.** 2015. Studies on the effect of bio-pesticides on muskmelon wilt (*Fusarium oxysporum* f.sp. *melonis*). *HortFlora Research Spectrum*, 4(3):250-254.
- Noling, J.W. and J.O. Becker.** 1994. The challenge of research and extension to define and implement alternatives to methyl bromide. *Journal of Nematology*, 26(4S):573-586.
- Okubara, P.A. and T.C. Paulitz.** 2005. Root defense responses to fungal pathogens: A molecular perspective. *Plant and Soil*, 274:215-226. <https://doi.org/10.1007/s11104-004-7328-9>
- Osman, E.A.M., S.E.M. El Nahas, M.M. Youssef and M.E.M. Hassan.** 2019. Use of organic acids for controlling damping-off caused by *Rhizoctonia solani* on cotton. *Journal of Plant Pathology (Mansoura University, Egypt)*, 10(4):205-210. <https://doi.org/10.21608/jppp.2019.40929>
- Oumouloud, A., M. El-Otmani, H. Chikh-Rouhou, A.G. Claver, R.G. Torres, R. Perl-Treves and J.M. Alvarez.** 2013. Breeding melon for resistance to Fusarium wilt: recent developments. *Euphytica*, 192(2):155-169. <https://doi.org/10.1007/s10681-013-0904-4>
- Paiva, N.A.M., N.J.D.A. Melo, M.M.Q. Ambrosio, G.H.S. Nunes and R.U.I Sales Junior.** 2022. Growth rate, pathogenicity and fungicide sensitivity of *Macrophomina* spp. From weeds melon and watermelon roots. *Revista Caatinga*, 35(3):537-547. <https://doi.org/10.1590/1983-21252022v35n304rc>
- Picó, B., C. Roig, A. Fita and F. Nuez.** 2008. Quantitative 441 detection of *Monosporascus cannonballus* in infected melon roots using real-time PCR. *European Journal of Plant Pathology*, 12:147-156.
- Porhanife, H.** 2010. Biological fertilizer. *Agriculture world* www.worldagronomy.blogfa.com
- Rajesh, C., K. Rajamanikkam, G.N.R. Vadivu and K. Palanichelvam.** 2019. Coelomic fluid of earthworm, *Eudrilus eugeniae*, inhibits the growth of fungal hyphae, in Vitro. *International Journal of Engineering and Advanced Technology*, 9(1S4):792-796. <https://doi.org/10.35940/ijeat.A1146.1291S419>
- Robinson R.W. and D.S. Deckers-Walters.** 1997. Cucurbits. Centre for Agriculture and Bioscience International, Wallingford, UK. 226 pp.
- Rocha, R.H.C., E.O. Sliva, L.C.C. Salomao and M.C. Ventrella.** 2010. Caracterização morfoanatómica do melão Gália no ponto de colheita. *Revista Brasileira de Fruticultura*, 32(2):375-385. <https://doi.org/10.1590/S0100-29452010005000048>
- Salari, M., N. Panjehkeh, Z. Nasirpoor and J. Abkhoo.** 2012. Reaction of melon (*Cucumis melo* L.) cultivars to soil-borne plant pathogenic fungi in Iran. *African Journal of Biotechnology*, 11(87):15324-15329.
- Savory, E.A, L.L. Granke, L.M. Quesada-Ocampo, M. Varbanova, M. K. Hausbeck and B. Day.** 2011. The cucurbit downy mildew pathogen *Pseudoperonospora cubensis*. *Molecular Plant Pathology*, 12(3):217-226. <https://doi.org/10.1111/j.1364-3703.2010.00670.x>
- Schlegel, I., N. T. Doan, N. de Chazal and G.D. Smith.** 1998. Antibiotic activity of new cyanobacterial isolates from Australia and Asia against green algae and cyanobacteria. *Journal of Applied Phycology*, 10(5):471-479. <https://doi.org/10.1023/A:1008042619686>
- Schoelz, J., D. Volenberg, M. Adhab, Z. Fang, V. Klassen, C. Spinka and M. Al Rwahnih.** 2021. A survey of viruses found in grapevine cultivars grown in Missouri. *American Journal of Enology and Viticulture*, 72(1):73-84. <https://doi.org/10.5344/ajev.2020.20043>
- Seong, Y.J., J. Hak and K. Yong.** 2017. Isolation and identification of N-butyl-tetrahydro-5-oxofuran-2-carboxamide produced by Bacillus sp. L60 and its antifungal activity. *Journal of Basic Microbiology*, 57(3):283-288. <https://doi.org/10.1002/jobm.201600481>
- Shafeek, M.R., A.M. Shaheen, E.H. Abd El-Samad, F.A. Rizk and F.S. Abd El-Al.** 2015. Response of growth, yield and fruit quality of cantaloupe plants (*Cucumis melo* L.) to organic and mineral fertilization. *Middle East Journal of Applied Science*, 5(1):76-82.
- Shalaby, M.E. and M.F. El-Nady.** 2008. Application of *Saccharomyces cerevisiae* as a biocontrol agent against Fusarium infection of sugar beet plants. *Acta Biologica Szegediensis*, 52(2), 271-275.
- Sharma, K. and V.K. Garg.** 2018. Vermicomposting: a green technology for organic waste management. Pp. 199-235. *In: Waste to Wealth. Energy, Environment, and Sustainability.* R. Singhaia, R. Agarwal, R. Kumar and R. Sukumaran (eds.) Springer, Singapore.
- Silva, S.G.A., M.M. Costa, A.M.S. Cardoso, L.V. Nascimento, K.A. Barroso, G.H.S. Nunes, L.H. Pfenning and M.M.Q. Ambrósio.** 2023. *Fusarium falciforme* and *Fusarium suttonianum* cause root rot of melon in Brazil. *Plant Pathology*, 72(4):721-730. <https://doi.org/10.1111/ppa.13701>
- Silvia S.M., P. Bagnaresi, S. Sestili, C. Biselli, A. Zechini, L. Orrù, L. Cattivelli and N. Ficcadenti.** 2017. Transcriptome analysis of the melon-*Fusarium oxysporum* f. sp. *melonis* race 1.2 patho system in susceptible and resistant plants. *Frontiers in Plant Science*. 8:362. <https://doi.org/10.3389/fpls.2017.00362>
- Sinha, R.K., D. Valani, K. Chauhan and S. Agarwal.** 2010. Embarking on a second green revolution for sustainable agriculture by vermiculture biotechnology

using earthworms: reviving the dreams of Sir Charles Darwin. *Journal of Agricultural Biotechnology and Sustainable Development*, 2(7):113-128.

- Smith-Becker, J., N.T. Keen and J.O. Becker.** 2003. Acibenzolar- S-methyl induces resistance to *Colletotrichum lagenarium* and cucumber mosaic virus in cantaloupe. *Crop Protection*, 22(5):769-774. [https://doi.org/10.1016/S0261-2194\(03\)00044-9](https://doi.org/10.1016/S0261-2194(03)00044-9)
- Sood, M., D. Kapoor, V. Kumar, M.S. Sheteiwiy, M. Ramakrishnan, M. Landi, F. Araniti and A. Sharma.** 2020. Trichoderma: the “secret” of a multitalented biocontrol agent. *Plants*, 9(6):762. <https://doi.org/10.3390/plants9060762>
- Stapleton, J.J.** 2000. Soil solarization in various agricultural production systems. *Crop Protection*, 19(8):837-841. [https://doi.org/10.1016/S0261-2194\(00\)00111-3](https://doi.org/10.1016/S0261-2194(00)00111-3)
- Stone, A.G., S.J. Scheuerell and H.M. Darby.** 2004. Suppression of soilborne diseases in field agricultural systems: organic matter management, cover cropping, and other cultural practices. Pp. 131-177. *In: Soil Organic Matter in Sustainable Agriculture* F. Magdoff

and R.R. Weil (eds.), CRC Press, Boca Raton, Fla., USA

- Sulaiman, I.S.C. and A. Mohamad.** 2020. The use of vermiwash and vermicompost extract in plant disease and pest control. Pp. 187-201. *In: Natural Remedies for Pest, Disease and Weed Control.* C. Egbuna and B. Sawicka. Academic Press: Cambridge, MA, USA.
- Williamson-Benavides, B.A. and A. Dhingra.** 2021. Understanding root rot disease in agricultural crops. *Horticulturae*, 7(2):33. <https://doi.org/10.3390/horticulturae7020033>
- Wu, H.S., Y.D. Liu, X.I. Yang, X.Q. Chen, Z.H. Wang, X.Y. Kong, X.X. Liu and S. Yan.** 2010. Growth responses of in vitro *Fusarium oxysporum* f. sp. *niveum* to external supply of tannic acid. *Journal of Environmental Biology*, 31(6):1017-1022.
- You, X., D. Wakana, K. Ishikawa, T. Hosoe and M. Tojo.** 2019. Antifungal activity of compounds isolated from bamboo vermicompost against *Rhizoctonia solani* AG1-IB. *Advances in Microbiology*, 9(12):957-970. <https://doi.org/10.4236/aim.2019.912061>

Received: August 24, 2023; Accepted: October 27, 2023

تاريخ الاستلام: 2023/8/24؛ تاريخ الموافقة على النشر: 2023/10/27